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# **1 Ideas and perspectives: A strategic assessment of methane and nitrous oxide measurements 2 in the marine environment**

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**Abstract.** In the current era of rapid climate change, accurate characterization of climate-relevant gas dynamics - namely production, consumption and net emissions - is required for all biomes, especially those ecosystems most susceptible to the impact of change. Marine environments include regions that act as net sources or sinks for a number of climate-active trace gases including methane (CH<sub>4</sub>) and nitrous oxide (N<sub>2</sub>O). The temporal and spatial distributions of CH<sub>4</sub> and N<sub>2</sub>O are controlled by the interaction of complex biogeochemical and physical processes. To evaluate and quantify the importance of these mechanisms relevant to marine CH<sub>4</sub> and N<sub>2</sub>O cycling requires a combination of traditional scientific disciplines including oceanography, microbiology, and numerical modeling. Fundamental to all of these efforts is ensuring that the datasets produced by independent scientists around the world are comparable and interoperable. Equally critical is transparent communication within the research community about the technical improvements required to increase our collective understanding of marine CH<sub>4</sub> and N<sub>2</sub>O. An Ocean Carbon & Biogeochemistry (OCB) sponsored workshop was organized to enhance dialogue and collaborations pertaining to marine CH<sub>4</sub> and N<sub>2</sub>O. Here, we summarize the outcomes from the workshop to describe the challenges and opportunities for near-future CH<sub>4</sub> and N<sub>2</sub>O research in the marine environment.



65

## 66 **1. Background**

67 The most abundant greenhouse gases in the troposphere, excluding water vapor, are carbon  
 68 dioxide (CO<sub>2</sub>), methane (CH<sub>4</sub>), and nitrous oxide (N<sub>2</sub>O). Together they account for more than  
 69 80% of the total radiative forcing (IPCC, 2013) and their current tropospheric mole fractions and  
 70 rates of increase are unprecedented in recent Earth history (Ciais et al., 2013; Burke et al., 2020;  
 71 Fig. 1a). While CO<sub>2</sub> is the most abundant of the three greenhouse gases, CH<sub>4</sub> and N<sub>2</sub>O both have  
 72 a higher warming potential than CO<sub>2</sub> (Montzka et al., 2011). To accurately constrain the  
 73 contribution of CH<sub>4</sub> and N<sub>2</sub>O to Earth's radiation budget requires their sources and sinks to be  
 74 quantified with high resolution at the global scale.

75 The oceans are a fundamental component of the global climate system and are a net source of  
 76 tropospheric CH<sub>4</sub> and N<sub>2</sub>O at the global scale, although local to regional budgets may include  
 77 both source and sink aspects. There are far fewer marine measurements of dissolved CH<sub>4</sub> and  
 78 N<sub>2</sub>O than of dissolved CO<sub>2</sub> and while there is substantial international coordination with regard  
 79 to CO<sub>2</sub> analysis, calibration and data reporting, no such coordination yet exists for CH<sub>4</sub> and N<sub>2</sub>O  
 80 (Wilson et al. 2018). Given the increasing prominence of climate change on scientific and  
 81 societal agendas, greater coordination among the marine CH<sub>4</sub> and N<sub>2</sub>O scientific community to  
 82 provide more targeted measurements and increase the quality and interoperability of CH<sub>4</sub> and  
 83 N<sub>2</sub>O observations is particularly timely.

84 Despite the lack of an international coordinating framework, there have been important  
 85 advances in our understanding of marine CH<sub>4</sub> and N<sub>2</sub>O in numerous research disciplines, ranging  
 86 from cellular metabolism and model microbial systems to large-scale modeling. For example,  
 87 recent work identified novel microorganisms and metabolic pathways in the production of N<sub>2</sub>O  
 88 (Trimmer et al., 2016; Caranto and Lancaster, 2017) and CH<sub>4</sub> (Repeta et al. 2016; Bižić et al.,  
 89 2020). Earth system models now incorporate improved N<sub>2</sub>O parameterizations to better resolve  
 90 the ocean's role in the global N<sub>2</sub>O cycle (Battaglia and Joos, 2018). New techniques enable the  
 91 discrimination of ancient and modern dissolved CH<sub>4</sub> (Sparrow et al., 2018) and the transfer of  
 92 CH<sub>4</sub>-derived carbon to other carbon pools (Pohlman et al., 2011; Garcia-Tigreros and Kessler,  
 93 2018). Other technological and analytical advances include improved near-continuous  
 94 spectroscopic analysis that yield greater sampling resolution in surface waters (e.g. Gülzow et



95 al., 2011; Arévalo-Martínez et al., 2013; Erler et al., 2015) and the deployment of analytical  
 96 devices on robotic vehicles (Nicholson et al., 2018).

97 These scientific advances and an improvement in the quantity and quality of CH<sub>4</sub> and N<sub>2</sub>O  
 98 observations are timely given that large areas of both the open and coastal ocean remain under-  
 99 sampled (Fig. 1b). This leads to uncertainty in oceanic CH<sub>4</sub> and N<sub>2</sub>O inventories, their rates of  
 100 production and consumption, and their emissions. This is problematic given that the marine  
 101 environment is susceptible to an accelerating rate of anthropogenic change that will continue to  
 102 modify the global cycles of carbon and nitrogen into the future. Environmental impacts on  
 103 marine CH<sub>4</sub> and N<sub>2</sub>O distributions include increasing seawater temperatures, decreasing  
 104 concentrations of dissolved oxygen (O<sub>2</sub>), acidification, retreat of ice and mobilization of carbon  
 105 substrates from former permafrost, altering coastal run-off, and eutrophication (IPCC, 2019).  
 106 These impacts will undoubtedly alter future CH<sub>4</sub> and N<sub>2</sub>O exchange with the atmosphere, but the  
 107 directions and magnitudes of these modified fluxes remains insufficiently understood.

108 A need to resolve the uncertainties prompted an evaluation of the collective ability of the  
 109 international scientific community to accurately determine the distribution and emissions of CH<sub>4</sub>  
 110 and N<sub>2</sub>O, and the physical-biogeochemical factors that determine them. This became the focus  
 111 of a marine CH<sub>4</sub> and N<sub>2</sub>O workshop hosted by the Ocean Carbon and Biogeochemistry (OCB)  
 112 program at Lake Arrowhead, California in October 2018. The workshop considered CH<sub>4</sub> and  
 113 N<sub>2</sub>O equally on the same agenda, even though nearly all field, laboratory, and modeling studies  
 114 examine these trace gases separately. The rationale for this dual approach is that CH<sub>4</sub> and N<sub>2</sub>O  
 115 share common considerations of the physical, chemical, and microbial processes that dictate their  
 116 water-column distributions (Bakker et al., 2014; Bodelier and Steenbergh, 2014). In addition,  
 117 many of the analytical procedures for quantifying CH<sub>4</sub> and N<sub>2</sub>O and the subsequent data quality  
 118 assurances share many common requirements. The opportunity to bring a large research  
 119 community together to increase dialogue and encourage the cross-fertilization of ideas was thus  
 120 considered very valuable. This article articulates the workshop outcomes framed in the context  
 121 of current marine CH<sub>4</sub> and N<sub>2</sub>O research and explores future research opportunities and  
 122 challenges.

123

## 124 2. Coordination of oceanic CH<sub>4</sub> and N<sub>2</sub>O measurements



Our understanding of the temporal and spatial distributions of oceanic CH<sub>4</sub> and N<sub>2</sub>O derives from over five decades of open ocean and coastal observations, including targeted expeditions, repeat hydrographic surveys, and time-series monitoring, each of which has been crucial to the development of our current knowledge (Fig. 2). Targeted programs have enabled invaluable insights into the role of oxygen deficient zones in N<sub>2</sub>O cycling (Babbin et al., 2015; Bourbonnais et al., 2017; Frey et al., 2020) and the exploration of CH<sub>4</sub>-rich seeps and vents (Foucher et al., 2009; Suess, 2010; Boetius and Wenzhöfer, 2013). Basin-scale repeat hydrographic surveys (e.g. the international GO-SHIP program) have facilitated extensive water-column mapping to identify relevant water masses and evaluate ventilation rates (e.g. de la Paz et al., 2017). Other oceanic surveys have focused exclusively on surface sampling, using continuous equilibrator systems connected to various gas analyzers to yield high-resolution surface concentration fields of CH<sub>4</sub> and N<sub>2</sub>O (Gülzow et al., 2013; Erler et al., 2015; Kodovska et al., 2016; Thornton et al., 2016a; Pohlman et al., 2017). In contrast, sustained long-term time-series measurements of CH<sub>4</sub> and N<sub>2</sub>O at fixed monitoring stations are relatively few, but they span a range of latitudes and biogeochemical provinces (Fig. 2). The time-series observations provide the contextual background for seasonal and interannual variation that allow long-term temporal trends and episodic events to be identified and evaluated (Farías et al., 2015; Wilson et al., 2017; Ma et al., 2019). Overall, the majority of measurements enable the variability in marine CH<sub>4</sub> and N<sub>2</sub>O to be quantified at the mesoscale or greater (i.e. from hundreds of kilometers to ocean basins), with monthly to annual resolution but there are substantially fewer datasets at the sub-mesoscale level (i.e. <10 km and hours to days) (Fig. 3). A major reason for the limited sampling at the sub-mesoscale level is that it necessitates high-resolution measurements to resolve the heterogeneous variability that exists at these time-space scales. Such analyses have only recently become technically feasible (discussed in more detail in Section 6).

Until recently there has been no formal coordination of observations across the CH<sub>4</sub> and N<sub>2</sub>O scientific community. In response to this, a Scientific Committee on Oceanic Research (SCOR) Working Group was initiated in 2014 entitled: ‘*Dissolved N<sub>2</sub>O and CH<sub>4</sub>: Working towards a global network of ocean time series measurements*’. A major goal of the SCOR Working Group was to unite the international community in joint activities conceived to improve and inform seagoing CH<sub>4</sub> and N<sub>2</sub>O analyses. An important activity was the preparation and distribution of common, combined gaseous CH<sub>4</sub> and N<sub>2</sub>O standards to twelve international laboratories, with



the aim of improving and standardizing calibration (Bullister et al., 2017). A subsequent inter-comparison of discrete seawater samples included the use of these standards and revealed the variability between laboratories. While there were some encouraging results from the intercomparison, such as the agreement between individual laboratories using contrasting techniques, overall a large range was observed in CH<sub>4</sub> and N<sub>2</sub>O concentration data generated by the participating laboratories (Wilson et al., 2018). Such analytical discrepancies weaken our collective ability as a community to evaluate temporal-spatial variability in marine CH<sub>4</sub> and N<sub>2</sub>O. The discrepancies also highlighted the need for Standard Operating Protocols (SOPs) for CH<sub>4</sub> and N<sub>2</sub>O analyses to facilitate standardization of sampling, measurement, and calibration, as well as the reporting of data and accompanying metadata in common repositories. The SOPs are currently in preparation with intended publication on the Ocean Best Practices network.

A data repository for oceanic CH<sub>4</sub> and N<sub>2</sub>O data known as the MarinE Methane and Nitrous Oxide database (MEMENTO) was established in 2009 (Bange et al., 2009; Kock and Bange, 2015). MEMENTO is now sufficiently mature to support descriptions of the broad-scale surface distributions of CH<sub>4</sub> and N<sub>2</sub>O (e.g. Suntharalingam et al., 2012; Zamora and Oschlies, 2014; Buitenhuis et al., 2018; Battaglia and Joos, 2018). Machine-learning mapping recently identified CH<sub>4</sub> and N<sub>2</sub>O distributions and various physical and biogeochemical predictor variables (e.g. depth, temperature, salinity, O<sub>2</sub>, nutrients, primary production) (Weber et al., 2019; Yang et al., 2020, Fig. 4). The application of gas transfer algorithms to the extrapolated oceanic CH<sub>4</sub> and N<sub>2</sub>O distributions helped decrease the uncertainty in estimates of global air-sea exchange fluxes (Fig. 4c), thereby fulfilling one of the key goals of MEMENTO (Bange et al., 2009). Net global open ocean emissions of N<sub>2</sub>O are now similarly estimated at 3–5 Tg N yr<sup>-1</sup> by both Yang et al. (2020) and the Global Nitrous Oxide Project (Tian et al., 2020). In comparison, net global ocean CH<sub>4</sub> emissions from machine-learning mapping were estimated at 6–12 Tg CH<sub>4</sub> yr<sup>-1</sup> (Weber et al., 2019), compared to 9–22 Tg CH<sub>4</sub> yr<sup>-1</sup> in the most up-to-date CH<sub>4</sub> synthesis (Saunois et al., 2020). However, the narrower range for machine-learning derived CH<sub>4</sub> emissions retains high uncertainty in regions such as the Arctic, where emissions are highly heterogeneous and compounded by seasonal ice cover. Identifying the causes for uncertainty in high emission regions will greatly aid future sampling campaigns, as is discussed in the following sections.

### 3. Methane in marine environments



187 In the surface waters of tropical and temperate oceans, the low supersaturation of CH<sub>4</sub> is  
 188 driven by aerobic production arising from the decomposition of methyl phosphonate in  
 189 phosphorus-depleted waters (Karl et al. 2008, Sosa et al., 2020), the degradation of methylated  
 190 sulfur compounds by phytoplankton (Klitzsch et al., 2019), and other processes (Schmale et al.,  
 191 2018). Deep within the ocean's pelagic interior, CH<sub>4</sub> is weakly undersaturated reflecting  
 192 depletion via microbial oxidation (Reeburgh 2007; Weber et al., 2019). Towards the coastline,  
 193 CH<sub>4</sub> supersaturation increases by orders of magnitude (Figure 5b), reflecting terrestrial inputs  
 194 (e.g. river and groundwater) and CH<sub>4</sub> diffusion and ebullition from shallow anoxic methane rich  
 195 sediments (Zhang et al., 2008; Borges et al., 2016; Upstill-Goddard and Barnes, 2016).  
 196 Supersaturation of CH<sub>4</sub> occurs frequently in the Arctic Ocean and its relatively shallow marginal  
 197 seas with the most extreme values observed in the Eurasian Arctic (e.g. Shakhova et al., 2010;  
 198 Damm et al., 2015; Kosmach et al., 2015; Thornton et al., 2016a; Fenwick et al., 2017).  
 199 Terrestrial and subsea permafrost are potential CH<sub>4</sub> sources to shelf waters in addition to CH<sub>4</sub>  
 200 hydrates that are found in marginal shelves globally (Ruppel and Kessler, 2017). Large point  
 201 source CH<sub>4</sub> emissions, such as seafloor gas seeps can be large sources to the atmosphere in small  
 202 localized areas (e.g. Thornton et al., 2020), but these sites remain particularly difficult to  
 203 parameterize in models. This reflects limited observations and a poor understanding of their  
 204 spatial distributions, the driving mechanisms, and the wider context within the carbon cycle. For  
 205 example, the upwelling of cold, nutrient-rich water that accompanies CH<sub>4</sub> ascending the water  
 206 column stimulates CO<sub>2</sub> consumption by photosynthesizing phytoplankton, rendering such CH<sub>4</sub>  
 207 seeps an overall net sink for climate-forcing gases (Pohlman et al., 2017). Recent work using  
 208 thermal infrared satellite retrievals indicates increased high-latitude oceanic CH<sub>4</sub> release in late  
 209 autumn, coincident with pycnocline breakdown and a deepening of the ocean mixed layer depth  
 210 thereby bringing deep CH<sub>4</sub> to the surface (Yurganov et al., 2019). This is especially notable in  
 211 the Kara and Barents Seas, but the remote observations have not yet been confirmed by surface  
 212 ocean measurements which are difficult and therefore rare, except during the Arctic summer.

213 Seabed CH<sub>4</sub> emissions are hypothesized to increase in a warming ocean through the  
 214 decomposition of gas hydrates, the degradation of subsea permafrost under some high-latitude  
 215 seas, and the increased biodegradation of sediment carbon (Romanovskii et al., 2005; Biastoch et  
 216 al., 2011; Ruppel and Kessler, 2017; Borges et al., 2019). Effort is thus focused on quantifying  
 217 the fraction of CH<sub>4</sub> generated in or released from marine sediments that ultimately enters the





218 atmosphere, particularly on shallow continental shelves and in coastal ecosystems. Natural  
 219 stable isotopes have been used to inform spatial and temporal changes in dissolved CH<sub>4</sub>  
 220 concentrations (e.g. Pack et al., 2011; Mau et al., 2012; Weinstein et al., 2016; Leonte et al.,  
 221 2017; Chan et al., 2019) and incubation experiments with added stable isotopes and radiotracers  
 222 have helped elucidate how oxidation (anaerobically in sediments and aerobically in the water  
 223 column), ebullition (where CH<sub>4</sub> pore water partial pressure exceeds sediment hydrostatic  
 224 pressure), and subsequent bubble dissolution in the water column interact to mitigate CH<sub>4</sub>  
 225 emissions to air (Steinle et al., 2015; Jordan et al., 2020). The information deriving from these  
 226 various approaches is inherently different but complementary. Isotope tracer incubations provide  
 227 snapshots of rates specific to the methanotrophic community and CH<sub>4</sub> concentration at the time  
 228 of sampling, whereas concentrations and isotopic gradients are used to infer *in situ* rates  
 229 integrated over space and time. A recent study deployed a remotely operated vehicle to examine  
 230 the isotopic fractionation of CH<sub>4</sub> during bubble ascent and used this to constrain the extent of  
 231 bubble dissolution (Leonte et al., 2018). This work demonstrated an experimental approach  
 232 established for broadly constraining water column CH<sub>4</sub> cycling directly from a surface research  
 233 vessel.

234 Despite the range of analytical and experimental approaches available, determining whether  
 235 the origin of the emitted CH<sub>4</sub> is seafloor release or aerobic production in the upper water column  
 236 remains problematic. To date there is no straightforward way to routinely distinguish between  
 237 seafloor derived and water column generated CH<sub>4</sub> for all locations. Even so, stable carbon and  
 238 hydrogen isotope measurements (i.e.  $\delta^{13}\text{C-CH}_4$  and  $\delta^2\text{H-CH}_4$ ) combined with ancillary data may  
 239 provide valuable source information. For example, combining these measurements with the ratio  
 240 of CH<sub>4</sub> to higher order hydrocarbons (e.g. ethene (C<sub>2</sub>H<sub>4</sub>) and ethane (C<sub>2</sub>H<sub>6</sub>)) can be used to infer  
 241 for example, whether the origin of the CH<sub>4</sub> is thermogenic, sub-seafloor, or biogenic within the  
 242 water column (Whiticar, 1999; Pohlman et al., 2009; Lan et al., 2019). Continuous shipboard  
 243 measurement of CH<sub>4</sub> isotopes in surface water (e.g. Pohlman et al., 2017) and in the atmospheric  
 244 boundary layer (Pankratova et al., 2019; Berchet et al., 2020) are now possible and they have  
 245 been used in combination with atmospheric inversion models to characterize and discriminate  
 246 marine-emitted CH<sub>4</sub> from other sources (Berchet et al., 2020). Application of this method to  
 247 land-based monitoring stations appears promising for apportioning CH<sub>4</sub> emissions from various  
 248 marine regions and sources (Thonat et al., 2019). Additionally, in regions where aerobic CH<sub>4</sub>



oxidation is substantial, the resulting isotopic fractionation generates measurable vertical and/or horizontal seawater gradients that can also be used to identify contrasting biogenic CH<sub>4</sub> sources (Leonte et al., 2020). However, the general overlap in isotope compositions of sediment CH<sub>4</sub> (e.g. Thornton et al., 2016b; Sapart et al., 2017) can complicate purely isotope-based determinations of sources.

Measurements of the natural radiocarbon content of dissolved oceanic CH<sub>4</sub>, while being highly specialized and requiring substantial amounts of ship time and processing (Kessler and Reeburgh, 2005; Sparrow and Kessler, 2017), provide valuable source information because the <sup>14</sup>C-CH<sub>4</sub> measurements are normalized to the same δ<sup>13</sup>C value and are unaffected by the extent of oxidation. The bubbles sampled from hydrate and active seafloor seeps are largely devoid of radiocarbon (Pohlman et al., 2009; Kessler et al., 2008; Douglas et al., 2016). However, CH<sub>4</sub> in sediments can also be derived from more modern or recently deposited organic material and an exact determination of individual contributions is hard to achieve (Kessler et al., 2008; Sparrow et al., 2018). The powerful insights made by radiocarbon-CH<sub>4</sub> investigations would be further strengthened by concurrent sampling of other analytes that offer CH<sub>4</sub> source information, such as clumped isotopes. Isotope clumping, the co-occurrence of two or more of the less abundant isotopes in a molecule (e.g. <sup>13</sup>C and <sup>2</sup>H or <sup>1</sup>H and <sup>2</sup>H), provides unique information on marine CH<sub>4</sub> sources (Stolper et al., 2014; Wang et al., 2015; Douglas et al., 2017; Young et al., 2017; Giunta et al. 2019). In this approach, the isotopic deviations in samples from their random probability distributions can give insight into formation temperature and the extent of biochemical disequilibrium. However, the sample size required for a clumped isotope analysis in the oceanic environment away from areas of seafloor emission is large and exceeds the already demanding volume requirements for <sup>14</sup>C analyses by 1–2 orders of magnitude (Douglas et al., 2017). While the requirement of large sample size and lengthy measurement time currently preclude their more widespread application, clumped isotope measurements offer future promise in refining our understanding of the processes of marine CH<sub>4</sub> production and consumption.

#### 4. Nitrous oxide in marine environments

The large-scale spatial distribution of N<sub>2</sub>O in the global ocean is reasonably well-established. The highest open ocean N<sub>2</sub>O values are in upwelling environments, where concentrations extend



up to micromolar levels (Arévalo-Martínez et al., 2015) and production rates can be as high as 120 nM d<sup>-1</sup> (Frey et al., 2020). The highly elevated N<sub>2</sub>O concentrations can be proximal to regions with some of the lowest recorded N<sub>2</sub>O concentrations, in the cores of O<sub>2</sub> deficient zones. This coexistence of the highest and lowest observed N<sub>2</sub>O concentrations over vertical distances of tens of meters make upwelling regions a focal point for N<sub>2</sub>O research, particularly since O<sub>2</sub> deficient ocean zones are increasing in size (Stramma et al., 2011). In contrast, in the surface waters of the expansive oligotrophic ocean gyres, N<sub>2</sub>O is weakly supersaturated (103-105%) with respect to atmospheric equilibrium (Weiss et al., 1992; Wilson et al., 2017, Charpentier et al., 2010). Nitrous oxide becomes more highly saturated in the surface waters of equatorial upwelling regions due to the upward advection of N<sub>2</sub>O-rich waters (Arévalo-Martínez et al., 2017). For the Arctic Ocean, the data indicate low net N<sub>2</sub>O emissions, with some areas acting as net N<sub>2</sub>O sources and others as N<sub>2</sub>O sinks (Fenwick et al., 2017, Zhang et al., 2015).

Several parameters control net N<sub>2</sub>O emissions from the ocean, including temperature, salinity, dissolved O<sub>2</sub>, apparent oxygen utilization (AOU), nutrients, and microbial community abundance and composition. A recent modeling study trained with just three of these variables (chlorophyll, O<sub>2</sub>, and AOU) accounted for 60% of the observed variability in oceanic N<sub>2</sub>O concentrations (Yang et al., 2020), highlighting the importance of N<sub>2</sub>O in productive upwelling systems. Correlations between N<sub>2</sub>O and environmental variables provide some insight into the factors controlling its distribution, but they provide no information about the microorganisms or metabolic pathways involved. Microbial production of N<sub>2</sub>O occurs during the metabolic processes of nitrification and denitrification (Stein and Yung, 2003). To determine which process dominates N<sub>2</sub>O production at any given location requires the application of multiple methodological approaches, ideally in parallel.

One of the most commonly used approaches is the incubation of discrete water samples under *in situ* conditions with stable isotope (<sup>15</sup>N) addition such as <sup>15</sup>N enriched NH<sub>4</sub><sup>+</sup>, NO<sub>2</sub><sup>-</sup> or NO<sub>2</sub><sup>-</sup>/NO<sub>3</sub><sup>-</sup> to measure N<sub>2</sub>O production rates from nitrification and denitrification, respectively (e.g. Ji et al., 2017). These approaches also provide insight into the microorganisms involved. For example, N<sub>2</sub>O resulting from archaeal NH<sub>4</sub><sup>+</sup> oxidation is mostly formed from a combination of NH<sub>4</sub><sup>+</sup> and another N compound (e.g. NO<sub>2</sub><sup>-</sup>) whereas bacteria produce N<sub>2</sub>O from NH<sub>4</sub><sup>+</sup> alone (Santoro et al., 2011, Stieglmeier et al., 2014; Carini et al. 2018; Lancaster et al., 2018; Frey et al. 2020). Unfortunately, as with all incubation-based approaches <sup>15</sup>N techniques are subject to



bottle artifacts, and the strong dependence of  $\text{N}_2\text{O}$  production and consumption on ambient  $\text{O}_2$  increases the potential for contamination during the collection and manipulation of anoxic deep seawaters. Incubation based rate measurements are also compromised by abiotic  $\text{N}_2\text{O}$  production via chemodenitrification, specifically the reduction of  $\text{NO}_2^-$  coupled to  $\text{Fe}^{2+}$  oxidation, as observed in high Fe environments (Ostrom et al., 2016; Buchwald et al., 2016; Wankel et al., 2017). These issues highlight the need for incubation techniques that mitigate the effect of experimental artifacts (Stewart et al., 2012).

In addition to isotope addition and incubation, natural abundance water-column measurements of  $\text{N}_2\text{O}$  concentrations, isotopes, and isotopomers yield valuable rate and process information. These measurements are free from experimental artifacts and can be used to integrate over appropriate temporal and spatial scales. For example, nitrification in sunlit waters has been inferred from  $\text{N}_2\text{O}$  distributions (Dore and Karl, 1996), and  $\text{N}_2\text{O}$  production close to the ocean surface is a large contributor to the uncertainty in oceanic  $\text{N}_2\text{O}$  emissions (Ward et al., 1982; Zamora and Oeschlies, 2014). Isotopomers are isomers having the same number of each isotope of each element but differing in their structural positions. Nitrous oxide isotopomers are increasingly used, sometimes in combination with box models, to estimate the rates of different  $\text{N}_2\text{O}$  production pathways, in the upwelling systems off southern Africa (Frame et al., 2014) and Peru (Bourbonnais et al., 2017). There is however some disagreement about whether isotopomer signatures are robust indicators of the formation pathway (Yoshida and Toyoda, 2000; Sutka et al., 2006) or whether there is fractionation during production (Schmidt et al., 2004; Casciotti et al., 2018). Greater clarity is therefore required in the use of  $\text{N}_2\text{O}$  isotopes and isotopomers to infer metabolic pathways of  $\text{N}_2\text{O}$  formation. Notwithstanding this issue, field measurements of  $\text{N}_2\text{O}$  isotopes and/or isotopomers have the potential to greatly increase current experimental capabilities and robustness (Yu et al., 2020). However, the development of spectroscopic gas analysis systems that have been so advantageous to  $\text{CH}_4$  research has been slower for  $\text{N}_2\text{O}$ . This is due to the higher costs and the increased complexity of the laser systems, although progress is being made to improve instrumental precision, and to decrease matrix effects and spectral interferences (e.g. Harris et al., 2019).

A better understanding of the microorganisms responsible for  $\text{N}_2\text{O}$  production and consumption is fundamental to deriving more accurate estimates of process rates. For example, the metabolic activity of ammonia oxidizing archaea can exceed that of ammonia oxidizing



bacteria in the ocean (Santoro et al., 2010; Löscher et al., 2012; Fuchsman et al., 2017). The differing sensitivities of these archaea and bacteria to dissolved  $O_2$  (Stahl and de la Torre, 2012; Hink et al., 2017) are a critical factor in evaluating the microbial response to changing environmental conditions, as shown for the terrestrial environment (Prosser et al., 2020). Therefore, to understand the impact of deoxygenation on oceanic  $N_2O$  emission requires a better understanding of both archaeal and bacterial metabolisms and their environmental niches. Field-based sequencing not only characterizes the community but can highlight potential metabolic pathways when they might not otherwise be inferred. For example, transcripts encoding for  $N_2O$  consumption (*nosZ*) have repeatedly been identified in the oxic water column, despite denitrification being an anaerobic metabolic process (Wyman et al., 2013; Sun et al., 2017). The transcription of *nosZ* has been also located in highly dynamic  $O_2$  permeable coastal sediments (Marchant et al., 2017). Denitrification under aerobic conditions is attributed to fluctuations in  $O_2$ ,  $NO_3^-$ , organic matter and other parameters that affect the availability of electron donors and acceptors which ultimately influences whether a coastal environment is a net source or sink of  $N_2O$ , as discussed in the next section.

### 5. $CH_4$ and $N_2O$ in shallow marine environments

Coastal and other shallow (<50 m) marine systems are globally relevant  $CH_4$  and  $N_2O$  source regions. However, their emission rates to the atmosphere are weakly constrained in comparison with the open ocean. Several factors contribute to the uncertainty, including the high diversity of coastal and shallow marine ecosystems and lack of consistency in adequately defining them, locally heterogeneous conditions causing strong spatial and temporal concentration gradients, highly uncertain spatial distribution of  $CH_4$  seeps, a bias towards studies in the northern hemisphere, and incomplete or sometimes inappropriate sampling strategies (Al-Haj and Fulweiler, 2020). Until these issues are resolved it will remain difficult to adequately define the contribution from shallow marine systems to global  $CH_4$  and  $N_2O$  budgets. An important illustration of this is reflected in the prevailing view that large geological sources (e.g. seeps, mud volcanoes, and hydrates) are the main contributors to marine  $CH_4$  emissions (Ciais et al., 2013). The most recent modeled estimate of global marine  $CH_4$  emissions ( $6\text{--}12\text{ Tg } CH_4\text{ yr}^{-1}$ ) reported that near-shore environments (depths of 0–50 m) contribute a large and highly uncertain diffusive flux (Weber et al., 2019). A study of coastal ecosystems, in this case defined as shelf,



373 estuarine, and tidally influenced rivers, estimated them to contribute 7 Tg CH<sub>4</sub> yr<sup>-1</sup> (Anderson et  
 374 al., 2010) while another estimated 1–7 Tg CH<sub>4</sub> yr<sup>-1</sup> for estuaries alone (Borges and Abril, 2011).  
 375 Similar uncertainties exist for N<sub>2</sub>O. Estimates of coastal N<sub>2</sub>O emissions (which include coastal,  
 376 estuarine, and riverine sources) range from 0.1–2.9 Tg N yr<sup>-1</sup> (Ciais et al., 2013), although a  
 377 recent review of N<sub>2</sub>O production across a range of estuarine habitats placed N<sub>2</sub>O fluxes at the  
 378 lower end of these estimates (0.17–0.95 Tg N yr<sup>-1</sup>) (Murray et al., 2015). Based on these data,  
 379 coastal systems account for around one third of total marine N<sub>2</sub>O emissions (Yang et al., 2020).

380 The direct quantification of CH<sub>4</sub> and N<sub>2</sub>O emissions from shallow coastal ecosystems has  
 381 historically involved using gas concentrations measured in discrete water and air samples  
 382 combined with a gas transfer velocity ( $k_w$ ). For the coastal and open ocean, the dominant driver  
 383 of gas exchange is wind speed (e.g. Nightingale et al., 2000; Wanninkhof, 2014) whereas in  
 384 nearshore, shallow water environments the interaction of water, depth, and tidal current speeds  
 385 may be a major contributor to near surface turbulence. Several  $k_w$  parameterizations are now in  
 386 use for coastal waters (e.g. Raymond and Cole 2001; Kremer et al., 2003; Zappa et al., 2003;  
 387 Borges and Abril, 2011; Ho et al. 2011; Rosentreter et al., 2017; Jeffrey et al., 2018) which  
 388 increases the uncertainties associated with CH<sub>4</sub> and N<sub>2</sub>O emissions. For example, a fivefold  
 389 variation in CH<sub>4</sub> emissions from a single system occurred when applying different  
 390 parameterizations to the measured gradients in CH<sub>4</sub> (Ferrón et al., 2007).

391 In order to constrain emissions over small areas continuous air-sea fluxes can be measured  
 392 using free-floating chambers (e.g. Bahlmann et al., 2015; Rosentreter et al., 2018; Yang et al.,  
 393 2018; Murray et al., 2020), but issues related to turbulence modification may still generate flux  
 394 artifacts (Upstill-Goddard, 2006). To overcome these problems in the future, a greater reliance  
 395 on direct and robust continuous techniques for air-sea flux measurement, such as eddy  
 396 covariance (e.g. Podgrajsek et al., 2016) that avoids any need for  $k_w$ , will be necessary.  
 397 Combining this with new analytical techniques such as cavity enhanced absorption spectroscopy  
 398 (CEAS) and non-dispersive infrared (NDIR) should continue to improve the quality of such  
 399 estimates (McDermitt et al., 2011; Nemitz et al., 2018; Maher et al., 2019). Indeed, eddy flux  
 400 towers aboard ships (Thornton et al., 2020) and in coastal locations (Yang et al., 2016; Gutiérrez-  
 401 Loza et al., 2019) are now being equipped with CH<sub>4</sub> instrumentation that enables the integration  
 402 of CH<sub>4</sub> fluxes over large areas. There are fewer N<sub>2</sub>O flux estimates made with CEAS and NDIR  
 403 and the implementation of N<sub>2</sub>O sensors on eddy flux towers remains limited. Recently, N<sub>2</sub>O



emissions from three major Eastern Boundary Upwelling Systems were quantified using atmospheric measurements from coastal monitoring stations highlighting their ability to attain multi-year time-series measurements (Ganesan et al., 2020).

Flux towers at fixed locations provide a stable instrument platform and facilitate the collection of ancillary data such as water-column depth, tidal motions (Rosentreter et al., 2018; Huang et al., 2019; Pfeiffer-Hebert et al., 2019), and other information relating to diel processes (Maher et al., 2016). Such data are important because for example, the magnitude of CH<sub>4</sub> and N<sub>2</sub>O fluxes vary over a diel period depending on the redox environment as a result of tidal effects and changes in inorganic N and O<sub>2</sub> availability (Seitzinger and Kroeze, 1998; Call et al., 2015; Vieillard and Fulweiler, 2014; Maher et al., 2015; Murray et al., 2015; Foster and Fulweiler, 2019). The magnitude of CH<sub>4</sub> and N<sub>2</sub>O fluxes also varies over longer temporal scales (seasonally to yearly) due to additional factors such as groundwater inputs, adjacent land-use, dissolved O<sub>2</sub>, organic matter content and quality, and macrofaunal distributions (Barnes and Upstill-Goddard, 2011; Upstill-Goddard and Barnes, 2016; Gelesh et al., 2016; Bonaglia et al., 2017; Borges et al., 2018; Wells et al., 2018; Ray et al., 2019; Al-Haj and Fulweiler, 2020; Reading et al., 2020). To determine the contributing factors and resolve the spatial distributions, mobile sampling platforms such as small vessels (Müller et al., 2016; Brase et al., 2017; Tait et al., 2017), and autonomous vehicles (Manning et al., 2019) are essential. Recent improvements in gas sensors and in technology such as sonar and ebullition sensors will further increase our ability to measure dynamic fluxes (Maher et al., 2019; Lohrberg et al., 2020). Improvements to the quality and quantity of CH<sub>4</sub> and N<sub>2</sub>O measurements in coastal systems will enable the development of iterative forecast models, further improving estimates of global coastal CH<sub>4</sub> and N<sub>2</sub>O fluxes.

## 6. Leveraging culture studies to further our ecosystem understanding

A more complete understanding of marine CH<sub>4</sub> and N<sub>2</sub>O necessitates closer integration between biogeochemistry, model requirements, and targeted microbiological studies involving both single microorganism isolates and enrichment cultures. Marine CH<sub>4</sub> and N<sub>2</sub>O budgets deriving from both ‘bottom-up’ (e.g. emissions inventories, ocean and terrestrial process models) and ‘top-down’ (e.g. inverse analyses of atmospheric trace-gas measurements) approaches would greatly benefit from more highly constrained metabolic processes. Specifically, this includes rates of





CH<sub>4</sub> or N<sub>2</sub>O production and consumption for key model microorganisms, and the kinetic parameters associated with these metabolic rates. Reliable inventories of key microbially mediated process rates will improve the robustness of Earth System models used for predicting climate-mediated changes to marine CH<sub>4</sub> and N<sub>2</sub>O emissions.

For N<sub>2</sub>O, laboratory studies quantifying microbial process rates, such as for nitrification and denitrification, are relatively few (e.g. Frame and Casciotti 2010; Santoro et al. 2011; Löscher et al. 2012; Ji et al. 2015; Qin et al., 2017). Consequently, models largely continue to use process rates optimized using water column concentrations of N<sub>2</sub>O, O<sub>2</sub>, and related nitrogen cycle quantities (e.g. Battaglia and Joos, 2018; Buitenhuis et al., 2018; Landolfi et al., 2017). Future model parameterizations for N<sub>2</sub>O will require information on the variability of microbial process yields derived from culture studies with controlled varying conditions of O<sub>2</sub> (Goreau et al. 1980, Frame and Casciotti 2010, Löscher et al. 2012; Ji et al., 2018), pH (Breider et al., 2019; Hopkins et al. 2020), temperature, and nutrients. Automated incubation systems have measured N<sub>2</sub>O production kinetics and yield as functions of the concentrations of O<sub>2</sub> and total ammonia nitrogen (Molstad et al., 2007; Hink et al., 2017). Quantifying the physiology of relevant microorganisms and connecting them to environmental characteristics will provide insights into why, for example, some shallow marine habitats act as N<sub>2</sub>O sinks while others are N<sub>2</sub>O sources, or how N<sub>2</sub>O is produced in well oxygenated open-ocean waters, as compared to oxygen deficient zones.

For CH<sub>4</sub>, a key requirement to relate *in situ* CH<sub>4</sub> production with transport to atmospheric emissions is our ability to accurately determine rates of CH<sub>4</sub> oxidation. Fundamental issues include the challenges of cultivating methanotrophs and of replicating environmental conditions such as pressure and the chemistry of CH<sub>4</sub> gas bubbles. The increased emphasis on CH<sub>4</sub> dynamics in shallow water environments highlighted in Section 5, must be supported by culture-based measurements of CH<sub>4</sub> oxidation that control for temperature, O<sub>2</sub> and other important variables. In comparison to CH<sub>4</sub> oxidation, culture-based studies are used increasingly to identify organisms capable of aerobic CH<sub>4</sub> production and their underlying metabolic pathways (Carini et al., 2014; Klintzsch et al; 2019; Bižić et al., 2020).

Specific cellular yields and consumption rates of CH<sub>4</sub> and N<sub>2</sub>O are not the sole objective of culturing experiments. Cultivation of microorganisms involved in CH<sub>4</sub> and N<sub>2</sub>O production and consumption provides vital information into the physiology, metabolism, and interactions of environmentally relevant clades. When combined with genomic approaches, insights can





therefore be gained into the diversity and global distribution of organisms involved in CH<sub>4</sub> and N<sub>2</sub>O cycling. For CH<sub>4</sub> some unexpected physiologies have been revealed (Ettwig et al., 2010; Haroon et al., 2013; Ettwig et al., 2016), which has directed research into sources and sinks of CH<sub>4</sub> in the natural environment. Similarly, our understanding of how and when ammonia oxidizers produce N<sub>2</sub>O has been facilitated by studies of cultured nitrifiers and detailed analysis of their biochemistry (Stahl and de la Torre, 2012; Caranto and Lancaster, 2017). Recent combinations of cultivation studies with environmental genomics, albeit largely for terrestrial systems, have revealed a variety of denitrifiers, many of which are only involved in specific denitrification steps (Ganesh et al., 2014; Lycus et al., 2017; Hallin et al., 2018; Marchant et al., 2018; Conthe et al., 2019).

476

## 477 **7. Outlook and priorities for marine CH<sub>4</sub> and N<sub>2</sub>O measurements**

This perspectives article has assessed the collective ability of the scientific community to determine the spatial variability of marine CH<sub>4</sub> and N<sub>2</sub>O distributions, the underlying mechanisms that determine this variability, and the resulting sea-to-air emissions. Shallow marine environments and oxygen deficient zones are widely recognized as deserving of greater attention because they have high CH<sub>4</sub> and N<sub>2</sub>O concentrations with inherently high uncertainties that complicate any assessment of their emissions to air (Bange et al., 1994; Bange et al., 1996; Bakker et al., 2014; James et al., 2016; Borges et al., 2016; Tian et al., 2020). Fortunately, recent technological advances that have increased our ability to conduct high-resolution measurements allow an optimistic outlook for making substantial progress in quantifying the CH<sub>4</sub> and N<sub>2</sub>O budgets of these ecosystems. Even so, the inherent complexity of shallow marine environments clearly warrants a strategically coordinated approach to optimize the value of future studies. Issues to consider include identifying the locations of complementary sampling sites, standardizing sampling strategies and techniques, and agreeing the use of common ancillary measurements that set the broad biogeochemical context (Bange et al., 2019). In contrast to the open ocean, measurement campaigns in shallow water environments are amenable to the use of eddy covariance flux towers, and they have the potential to lever resources from existing observation networks, which in North America include the Long-Term Ecological Research network (LTER) and the National Estuarine Research Reserve (NERR) System (Novick et al., 2018). Indeed, such activities are already underway; an increasing number of flux towers are



497 being equipped for CH<sub>4</sub> measurements (Torn et al., 2019) and future efforts should focus on the  
498 inclusion of N<sub>2</sub>O (see Section 5).

499 We are encouraged that the Global Carbon Project with its objective of developing a  
500 complete picture of the global carbon cycle including interactions and feedbacks has expanded to  
501 include CH<sub>4</sub> (Saunio et al., 2020) and is now incorporating N<sub>2</sub>O (Tian et al., 2020). These  
502 Projects compile the most recent data from peer-reviewed analyses of the sources and sinks of  
503 atmospheric CH<sub>4</sub> and N<sub>2</sub>O from both natural and human activities. For example, the aquatic  
504 components of the recent Global Carbon Project N<sub>2</sub>O budget reported emissions from the open  
505 ocean, inland waters, estuaries and coastal zones. Low-oxygen oceanic regions associated with  
506 eastern-boundary upwelling zones, and the coastal ocean were identified as key regions with  
507 significant N<sub>2</sub>O variability requiring more detailed assessment via measurement campaigns and  
508 model analyses (Tian et al., 2020). Coordinating with global initiatives such as the Global  
509 Carbon Project and identifying other areas of synergistic CH<sub>4</sub> and N<sub>2</sub>O research of mutual  
510 benefit to oceanographers and scientists studying other biomes serve to strengthen the scientific  
511 achievements of all involved (Ganesan et al., 2019). Furthermore, as highlighted in Section 6,  
512 field observations alone are insufficient to improve the robustness of Earth System models and  
513 leveraging laboratory-based microbial process studies is highly recommended.

514 The success of any coordinated CH<sub>4</sub> and N<sub>2</sub>O research program relies heavily on having  
515 uniformly high confidence in the various resulting datasets and their interoperability, and we  
516 identify three key initiatives that are paramount to ensuring this:

517 (i) The first is to develop and adopt Standard Operating Protocols (SOPs) to help obtain  
518 intercomparable CH<sub>4</sub> and N<sub>2</sub>O datasets of the highest possible accuracy and precision. In our  
519 recent marine CH<sub>4</sub> and N<sub>2</sub>O inter comparison exercise we concluded that the diversity of  
520 analytical procedures employed by the participants was a major cause of high variability between  
521 the reported concentrations, highlighting an urgent requirement for CH<sub>4</sub> and N<sub>2</sub>O SOPs (Wilson  
522 et al., 2018). Consequently, these SOPs are now being compiled, and they will be freely  
523 available via the Ocean Best Practices System.

524 (ii) The second is the regular, routine inter comparison of measurements, by periodically  
525 distributing to the community “consensus material”, i.e. water samples in which CH<sub>4</sub> and N<sub>2</sub>O  
526 concentrations are known with high confidence, obtained by pooling analyses from several  
527 laboratories with demonstrated analytical capability. These will allow us to routinely monitor



528 data inter comparability and accuracy, particularly in the case of highly elevated concentrations  
 529 of CH<sub>4</sub> and N<sub>2</sub>O, i.e. those exceeding atmospheric equilibrium concentrations by at least an order  
 530 of magnitude.

531 (iii) The third activity is increased use and support for MEMENTO. Until now the main  
 532 function of MEMENTO has been as a data repository. In this regard, it has been very valuable in  
 533 supporting the modeling components of CH<sub>4</sub> and N<sub>2</sub>O research (see Section 3). We encourage a  
 534 much more widespread, routine, use of this data facility, with submitted data produced according  
 535 to the SOPs and inter comparison procedures. To maintain its relevance, MEMENTO must  
 536 continue to build its activities and develop into an ‘ocean CH<sub>4</sub> and N<sub>2</sub>O Atlas’. The international  
 537 marine carbon science community has widely embraced such an approach for CO<sub>2</sub>, by  
 538 submitting data to the Surface Ocean CO<sub>2</sub> Atlas (SOCAT), which was initiated in response to the  
 539 need for a quality controlled, publicly available, global surface CO<sub>2</sub> dataset (e.g. Bakker et al.,  
 540 2016). We believe establishing a similar data product for marine CH<sub>4</sub> and N<sub>2</sub>O to be essential  
 541 for supporting future global modeling efforts and to enhance and reward community  
 542 engagement.

543 The benefits of pursuing the three activities described above have already been clearly  
 544 demonstrated for carbon system measurements in the ocean. The intercomparability and high  
 545 accuracy and precision of carbon system measurements was achieved by streamlining  
 546 methodological approaches, universally adopting agreed SOPs, production of reference material,  
 547 and following community-driven quality control procedures (Dickson et al., 2007, Dickson et al,  
 548 2010). It is encouraging to see the marine CH<sub>4</sub> and N<sub>2</sub>O community beginning to move in a  
 549 similar direction.

550

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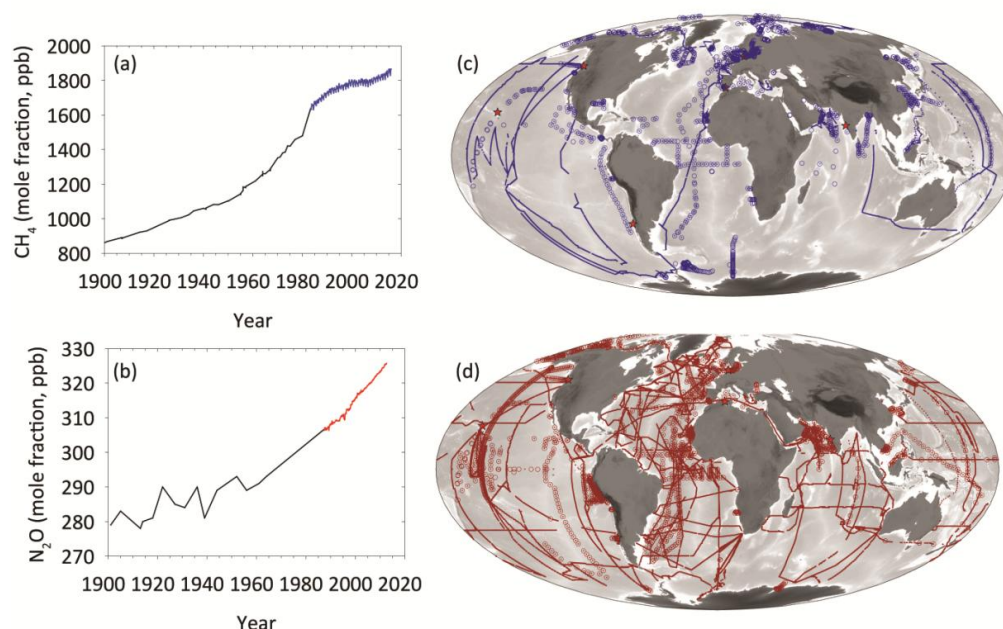


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1175 Figure 1. Atmospheric values of (a) CH<sub>4</sub> and (b) N<sub>2</sub>O with the black lines reconstructed from  
 1176 ice-core measurements (Etheridge et al., 1998; Machida et al., 1995) and the colored lines from  
 1177 Mauna Loa Observatory (<https://www.esrl.noaa.gov/gmd/dv/data/>). Global maps of marine (c)  
 1178 CH<sub>4</sub> and (d) N<sub>2</sub>O measurements available from the MEMENTO database  
 1179 (<https://memento.geomar.de/>). The 2018 workshop focused on the marine contribution to  
 1180 atmospheric CH<sub>4</sub> and N<sub>2</sub>O and the underlying microbial and biogeochemical control  
 1181 mechanisms.

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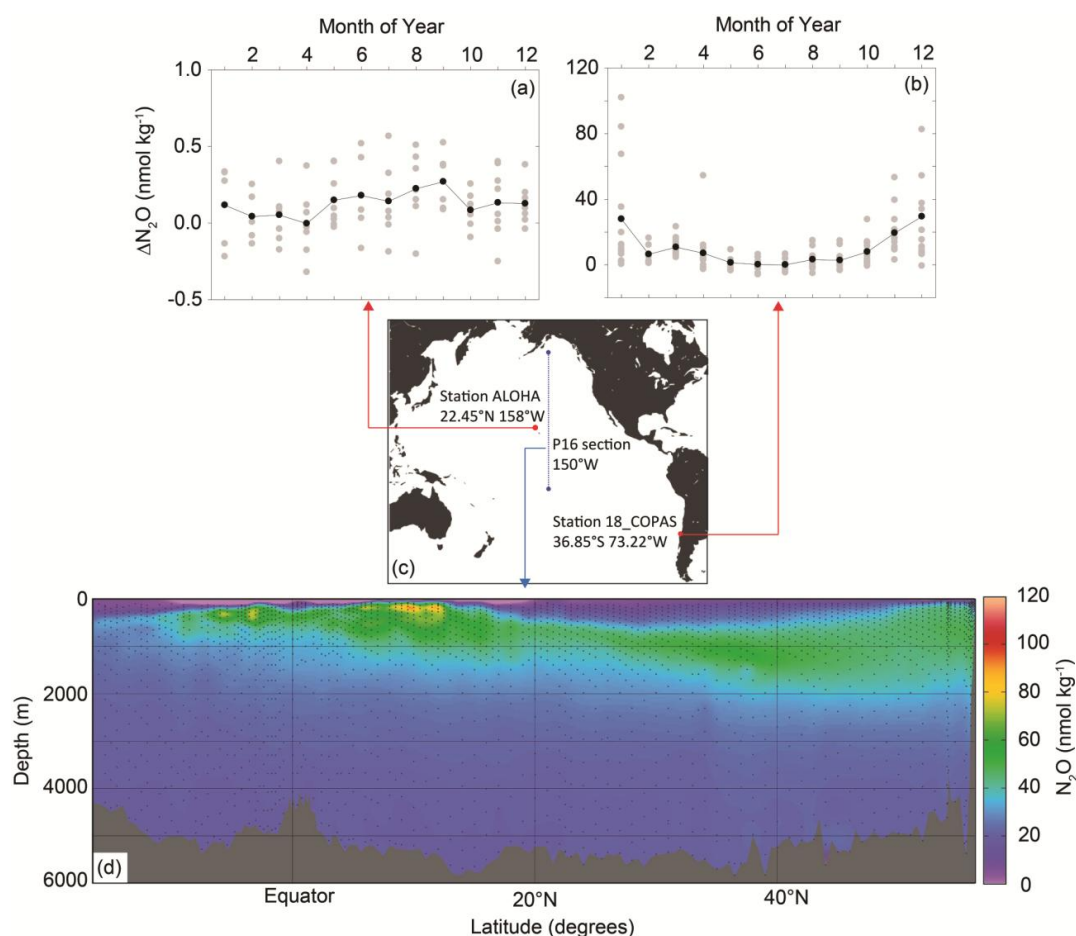


Figure 2. Repeat oceanic observations include both (a, b) fixed location time-series monitoring observations and (c,d) hydrographic surveys. Together, such field observation programs helps resolve temporal variability ranging from months to years and spatial variability at the ocean basin scale (see Fig. 3). The Station ALOHA data derive from Wilson et al. (2018), the Station 18 data derive from Farías et al. (2015), and the P16 transect was conducted in 2015 by the NOAA PMEL Tracer Group as part of the GO-SHIP program. The data shown in the plots are  $N_2O$  concentrations, either as  $\Delta N_2O$  (i.e. deviation from equilibrium value) or absolute values.

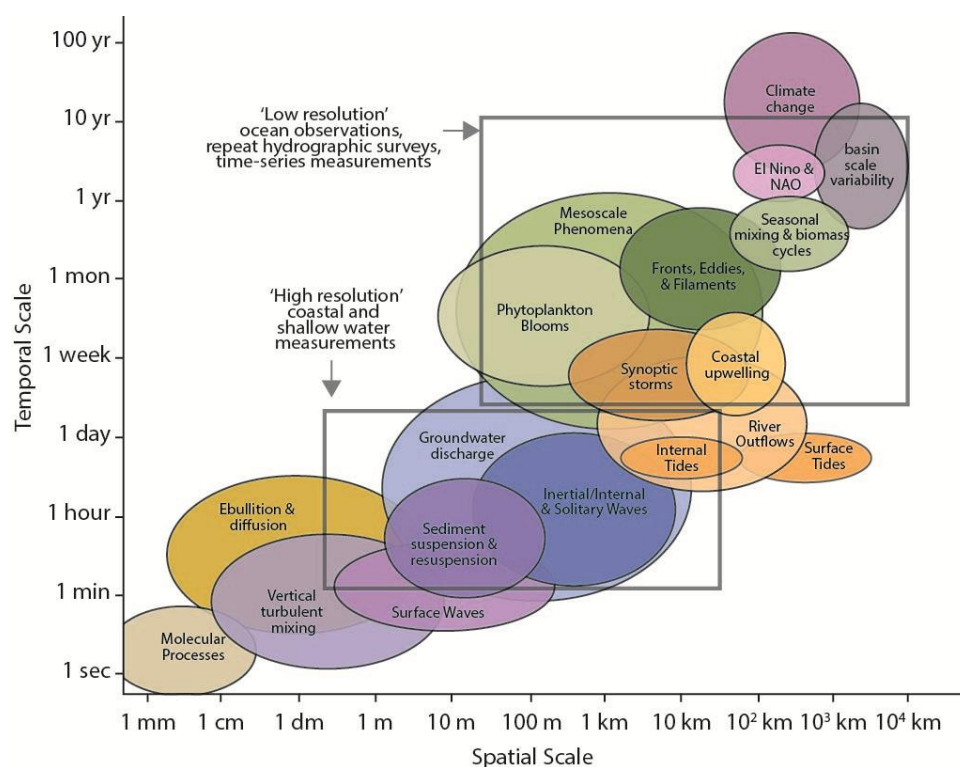


Figure 3. Time-space scale diagram illustrating various physical, biological, and climatological processes relevant to marine CH<sub>4</sub> and N<sub>2</sub>O (adapted from Dickey, 2003). To date, the majority of marine CH<sub>4</sub> and N<sub>2</sub>O measurements resolve variability at the mesoscale level or higher. Recent technological developments and the need to resolve concentrations and fluxes in shallow water environments will increase the number of measurements conducted at the sub mesoscale.

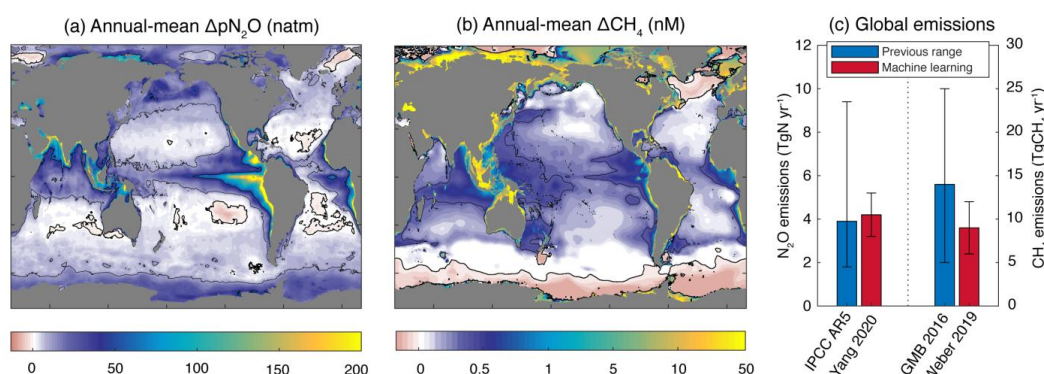


Figure 4. Distributions and emissions of marine  $CH_4$  and  $N_2O$ , (a) Air-sea  $N_2O$  disequilibrium mapped using a Regression Forest model (adapted from Yang et al., 2020), (b) Air-sea  $CH_4$  disequilibrium mapped using an Artificial Neural Network model (adapted from Weber et al., 2019). For consistency with the original publications, the air-sea disequilibrium is shown in different units for  $N_2O$  (partial pressure) and  $CH_4$  (concentration). (c) A summary of global ocean  $CH_4$  and  $N_2O$  emissions estimated by Yang et al. (2020) and Weber et al. (2019), compared to the estimates of the IPCC 5th Annual Report (IPCC AR5) and the Global Methane Budget (Saunois et al., 2016).

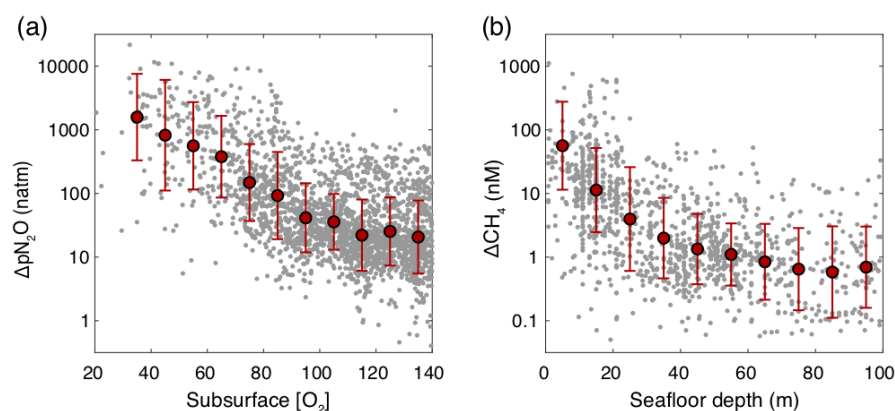


Figure 5. Key environmental predictors of surface ocean  $\text{CH}_4$  and  $\text{N}_2\text{O}$  gradients. (a) Excess air-sea  $\text{N}_2\text{O}$  is best predicted by  $\text{O}_2$  concentrations in the subsurface water-column (base of the mixed layer to a depth of 100 m) (adapted from Yang et al., 2020). (b) Excess  $\text{CH}_4$  is best predicted by seafloor depth, reflecting the supply from anoxic sediments (adapted from Weber et al., 2019). The grey dots represent individual data points and the red dots with error bars represent mean  $\pm 1$  s.d. of binned data, using  $\text{O}_2$  bins of  $10 \mu\text{M}$  width and seafloor depth bins of 10 m width.