

Phylogenetics of wigeons and allies (Anatidae: *Anas*): the importance of sampling multiple loci and multiple individuals

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Abstract

Species-level DNA phylogenies frequently suffer from two shortcomings—gene trees usually are constructed from a single locus, and often species are represented by only one individual. To evaluate the effect of these two shortcomings, we tested phylogenetic hypotheses within the wigeons and allies, a clade of *Anas* ducks (Anatidae) composed of five species. We sequenced two nuclear introns from the Z-chromosome-linked chromo-helicase binding protein gene (CHD1Zb and CHD1Za) and the mitochondrial DNA (mtDNA) control region for multiple individuals sampled from widespread geographic locations. We compared these phylogenies to previously published phylogenies constructed from morphology and protein coding regions of mtDNA. Relative to other nuclear introns, CHD showed remarkable phylogenetic utility. Of the 26 CHD1Zb alleles identified, only one was shared between two species, and the combined CHD datasets revealed that four of the five species were consistent with monophyly. Several species shared mtDNA haplotypes, which probably was a result of interspecific hybridization. Overall, the nuclear CHD tree and the mtDNA tree were more congruent with coding regions of mtDNA than they were with morphology.

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1. Introduction

Constructing phylogenetic trees from DNA sequences has become standard practice, but many molecular phylogenetic analyses suffer from two shortcomings. First, the vast majority of animal species-level studies have relied solely on mitochondrial DNA (mtDNA). Although high mutation rates, low effective population size, and lack of recombination make

mtDNA an excellent marker (Avice et al., 1987; Moore, 1995; Moritz et al., 1987), mtDNA is a single linkage group that can be susceptible to stochastic lineage sorting and hybridization, either of which can mislead phylogenetic inferences (Avice, 2000; Ballard and Whitlock, 2004; Funk and Omland, 2003; Palumbi et al., 2001). A second shortcoming of many species-level phylogenies is that only one or a few representatives of each taxon are sampled (Funk and Omland, 2003; Omland, 1997). In a review of mtDNA phylogenetic and phylogeographic studies, Funk and Omland (2003) found that 17% of 331 bird species had species-level paraphyly or polyphyly; some individuals of a species had mtDNA that was more

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similar to other species than to conspecifics. Hence, phylogenetic relationships among closely related species are often more complex than can be suggested by the inclusion of a single representative (also see Omland et al., 1999). To obtain a better understanding of species-level phylogenetics, studies should evaluate multiple genetic loci (Hare, 2001; Pritchko and Moore, 1997) and attempt to include multiple individuals of each species from widespread geographical locations (Funk and Omland, 2003).

Nuclear introns provide independent characters for phylogenetic analyses, and several studies have found these markers to be useful at resolving deep phylogenetic relationships, such as at the genus and familial levels (e.g., Armstrong et al., 2001; Barker, 2004; Johnson and Clayton, 2000b; Shapiro and Dumbacher, 2001). However, the ability of introns to resolve relationships among closely related taxa has shown less promise (e.g., Allen and Omland, 2003; DeBry and Seshadri, 2001; Johnson and Clayton, 2000a). Nuclear introns accumulate substitutions at slower rates than mtDNA (Brown et al., 1979) and often do not offer sufficient resolving power to determine the relationships among closely related species. Even if nuclear introns contain enough substitutions to give phylogenetic signal, their slower lineage sorting rate, relative to mtDNA (Moore, 1995; Palumbi et al., 2001), increases the likelihood that species will not have sorted to monophyly (all intraspecific alleles sharing a more recent common ancestor with each other than with heterospecific alleles). However, Z-chromosome-linked loci may have higher phylogenetic utility than autosomal loci. First, Z-linked loci have three-quarters the effective population size of autosomal loci (males carry two Z-chromosomes, whereas females carry one), and therefore, they are expected to sort slightly more rapidly. In addition, Z-linked loci should undergo recombination only two-thirds as often as autosomal loci, because one-third of gametes for Z-linked loci segregate in the heterogametic sex where recombination presumably does not occur. Furthermore, mutation rates tend to be higher within the male genome than the female genome (e.g., Ellegren and Fridolfsson, 1997; Fridolfsson and Ellegren, 2000; Kahn and Quinn, 1999; Montell et al., 2001). Hence, Z-linked loci should accumulate mutations slightly more rapidly than autosomal loci. Finally, by sequencing females, one can avoid scoring alleles occurring in heterozygotes. CHD is a sex-linked gene, present on both the W- and Z-chromosomes, that codes for a chromo-helicase DNA binding protein (Ellegren and Fridolfsson, 1997; Fridolfsson and Ellegren, 1999). W- and Z-copies are highly divergent and chromosome-specific primers can be used to preferentially amplify the Z-copy. Pitra et al. (2002) sequenced CHD introns for phylogenetic analyses, but they did not provide a CHD tree. In this study, we further investigate the utility of CHD for phylogenetic studies.

Ducks of the genus *Anas* provide an excellent model system for evaluating multiple loci and the importance of sampling multiple individuals for testing phylogenetic hypotheses. Phylogenetic analyses have been conducted for the majority of *Anas* species, as well as several closely related genera, using morphology (Livezey, 1991) and mtDNA ND2 and cytochrome *b* sequences (Johnson and Sorenson, 1998, 1999). Analyses also have been conducted using a variety of additional character sets, although taxon sampling was less intensive in these studies (Donne-Gousse et al., 2002; Kessler and Avise, 1984; Patton and Avise, 1986; Tuohy et al., 1992). In many aspects, trees constructed from different sets of characters are similar (see Omland, 1994), but Johnson and Sorenson (1999) found significant incongruence between the phylogenies of Livezey (1991) and Johnson and Sorenson (1998). Hence, analyses of additional independent characters are needed to test these phylogenetic hypotheses.

Within *Anas* ducks, sampling mtDNA from one individual may not be representative of the species. This genus underwent a rapid radiation in the past few million years (Johnson and Sorenson, 1999), and there may have been an insufficient amount of time for mtDNA to have sorted to reciprocal monophyly for several closely related species (Avise et al., 1990; Johnson and Sorenson, 1999; McCracken et al., 2001; Omland, 1997). In addition, several species of *Anas* ducks have widespread geographical distributions that extend across multiple continents. Such widely distributed species may harbor cryptic lineages that are not revealed by sampling a single individual (e.g., Avise et al., 1992; Omland et al., 2000; Talbot and Shields, 1996; Zink et al., 1995). Finally, dabbling ducks are well known for their capacity to hybridize, and such hybridization often produces fertile offspring (Johnsgard, 1960; Tubaro and Lijtmaer, 2002; see also Rhymer and Simberloff, 1996). Interspecific hybridization may explain the polyphyletic relationships observed in some groups of ducks (e.g., mallards; Kulikova et al., 2004; McCracken et al., 2001; Rhymer et al., 1994), and hence can lead to incorrect inferences if a single individual is sampled. Overall, these characteristics of *Anas* ducks increase the likelihood of species-level paraphyly and polyphyly and the probability that a single individual will not be representative of the species.

In this study, we focus on the wigeons and their allies, a monophyletic clade of *Anas* ducks that consists of five species that are approximately 0.5–4.7% divergent based on mtDNA cytochrome *b* and ND2 sequences (see Johnson and Sorenson, 1998). The wigeons and allies [Livezey's (1991) genus *Mareca*, Johnson and Sorenson's (1999) "wigeon clade"] include the three wigeon species *Anas penelope* (Eurasian wigeon), *A. americana* (American wigeon), and *A. sibilatrix* (Chiloe wigeon), as well as *A. strepera* (gadwall) and *A. falcata* (falcated duck). Morphology and mtDNA agree in grouping these five

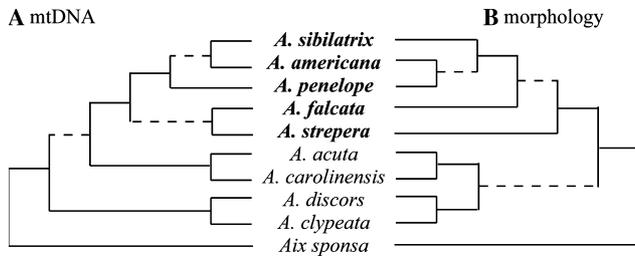


Fig. 1. Phylogenetic relationships among a subset of *Anas* ducks based on: (A) mtDNA ND2 and cytochrome *b* sequences (Johnson and Sorenson, 1998, 1999) and (B) morphology (Livezey, 1991). The wigeons and allies are shown in bold type; dashed lines connect the three nodes that disagree between mtDNA and morphology.

species as each other's closest relatives, but the relationships inferred by mtDNA and morphology differ within the clade (Fig. 1; Johnson and Sorenson, 1998, 1999; Livezey, 1991). Morphology supports a sister relationship between *A. penelope* and *A. americana*, placing *A. sibilatrix* in a basal position. However, mtDNA suggests that the divergence of *A. penelope* predated the divergence of *A. americana* and *A. sibilatrix*. Furthermore, morphology supports a sister relationship between *A. falcata* and the three wigeon species, with *A. strepera* basal to these four species, but mtDNA supports a sister relationship between *A. strepera* and *A. falcata* to the exclusion of the wigeons.

In addition to the within-clade incongruence between morphology and mtDNA, there also is disagreement in the placement of the wigeons and allies within the *Anas* phylogeny. Morphology suggests a sister relationship between this group and all other *Anas* ducks, which led Livezey (1991) to suggest elevating the wigeons and allies to genus *Mareca*. However, mtDNA suggests a sister relationship between the wigeons and allies and most other *Anas* ducks (Fig. 1B), including pintails, mallards, green-winged ducks, and Australasian teals, but to the exclusion of the blue-winged ducks (Johnson and Sorenson, 1998, 1999). Hence, Johnson and Sorenson's (1998, 1999) data clearly place the wigeons and allies as nested within *Anas* (Fig. 1A). In this study, we further evaluate the phylogenetic relationships within the wigeons and allies by sequencing sex-linked CHD nuclear introns and the mtDNA control region, and by sampling multiple individuals from widespread geographic locations. We also evaluate the placement of this group within *Anas* ducks using CHD.

2. Materials and methods

2.1. Sample collection

Each of the five wigeons and allies was densely sampled ($n=10$ – 22 individuals for each species), and sam-

pling was geographically widespread for all species, except *A. falcata* (Fig. 2). Muscle tissue was provided by museums or collected at the United States and the Canadian hunter wing surveys, feathers were salvaged from dead birds, and foot-webbing and blood were collected from captured birds (Appendix). There was no indication that nuclear pseudogenes of mitochondrial DNA origin (numts, Sorenson and Fleischer, 1996) were amplified from the blood. All ducks sampled were from the wild, except one *A. falcata* and one *A. sibilatrix* from captivity. We also sampled one individual each of *A. acuta* (Northern pintail), *A. crecca carolinensis* (North American green-winged teal), *A. clypeata* (Northern shoveler), *A. discors* (blue-winged teal), and *Aix sponsa* (North American wood duck). We retrieved homologous ingroup and outgroup mtDNA control region sequences from GenBank, including *A. strepera* (AY112944), *A. sibilatrix* (AY112944), *A. sibilatrix* (AY112943), *A. platyrhynchos* (mallard; AY112938), *A. acuta* (Northern pintail; AY112939), and *A. crecca crecca* (Eurasian common teal; AY112942) (Donne-Gousse et al., 2002).

2.2. Laboratory methods

DNA was extracted from samples using the DNeasy Tissue Kit (Qiagen, Valencia, CA, USA). We amplified 270–271 bp of a nuclear intron and 65 bp of flanking exon from the Z-chromosome CHD gene (intron b, CHD1Zb; Ellegren and Fridolfsson, 1997). PCR amplification was conducted using 1 μ M each of primers 3007F and 3112R, that specifically targeted the Z-chromosome (Ellegren and Fridolfsson, 1997), 1.5 U *Taq* DNA polymerase, 2.5 mM $MgCl_2$, and 0.25 mM each dNTP in 50 μ l total volume. PCR included an initial denaturing step of 94 $^{\circ}C$ for 9 min, followed by 35 cycles of 94 $^{\circ}C$ for 1 min, 54 $^{\circ}C$ for 1 min, and 72 $^{\circ}C$ for 1 min, and a final elongation step at 72 $^{\circ}C$ for 4 min. We cleaned PCR products with a QIAquick Kit (Qiagen, Valencia, CA) and sequenced both strands using the same primers and BigDye Terminator Cycle Sequencing Kits (Applied Biosystems, Foster City, CA). Excess dye terminators were removed using ethanol precipitation, and products were sequenced with an ABI 3100 automated sequencer (Applied Biosystems, Foster City, CA).

Sequences were aligned and edited using Sequencher 4.1 (Genecodes, Ann Arbor, MI). Because females are the heterogametic sex, they contained no polymorphic sites in CHD. However, several males were heterozygous at two or more positions. To determine the gametic phase of each allele, we used allele-specific priming (Bottema et al., 1993; see also Hare and Palumbi, 1999). In short, we designed primers that were identical at all sites except the last position on the 3'-end, which matched one of the polymorphic sites in the template sequence

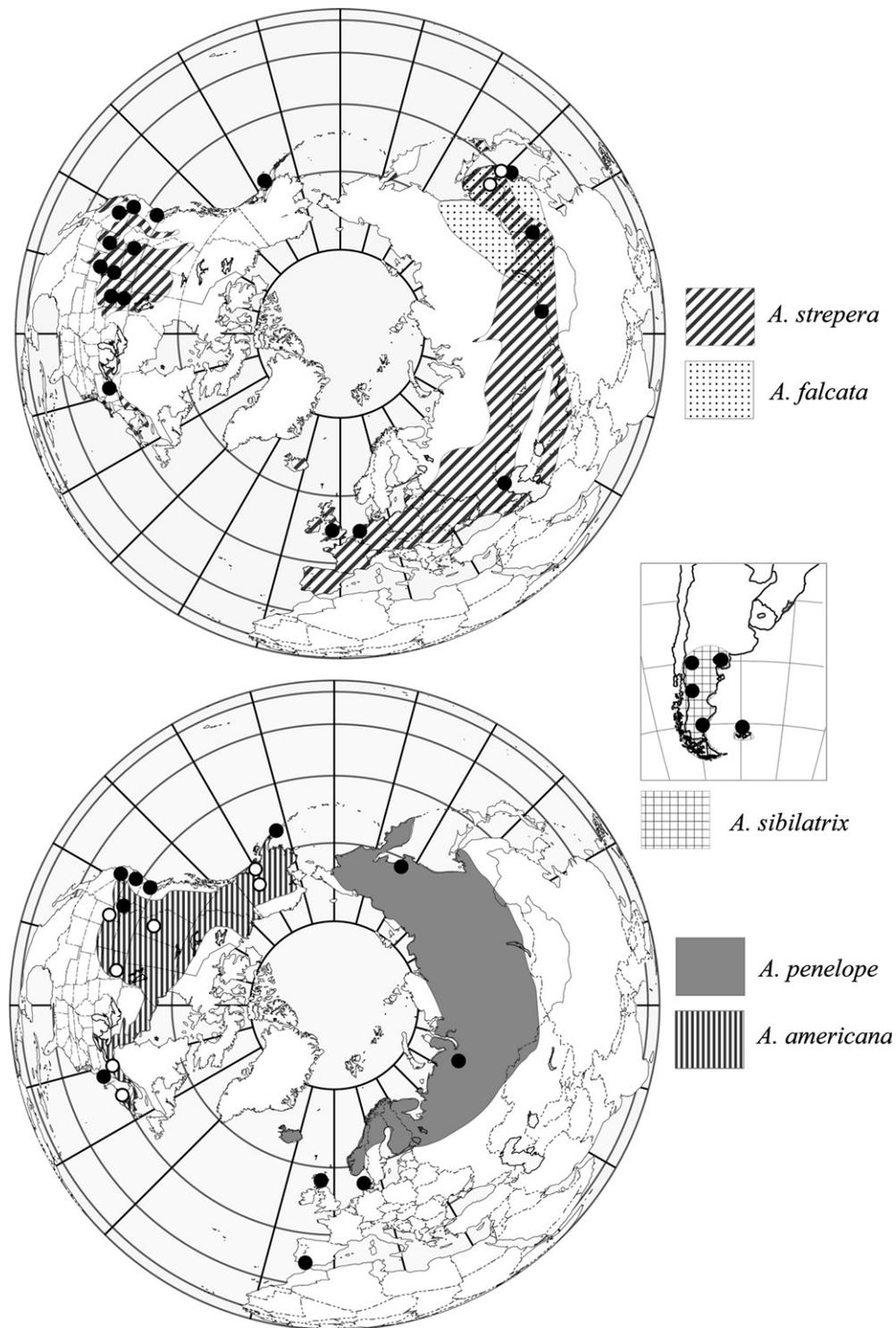


Fig. 2. Approximate breeding distributions of the five wigeons and allies (modified from LeSchack et al., 1997; Madge and Burn, 1988; Mowbray, 1999). Closed circles indicate sampling locations for *A. strepera*, *A. sibilatrix*, and *A. penelope*. Open circles indicate sampling locations for *A. falcata* and *A. americana*. For simplicity, only breeding distributions are illustrated, although all five species are seasonal migrants, and several individuals were sampled from migrating and over-wintering populations (see Appendix).

(Table 1). Thus, we were able to amplify and sequence both alleles individually for heterozygous males, using the PCR and sequencing protocols described above and primer-specific annealing temperatures (Table 1). One *A.*

penelope and one *A. falcata* were not polymorphic at the primer-specific position, and for these males, we used the technique of allele subtraction to determine the gametic phase (Clark, 1990).

Table 1
Primers used for Z-linked CHD introns

	Primer sequence	Annealing temperature (°C)
Allele-specific primers		
<i>A. penelope</i>		
3007pe.F	5'-ACACACAGGTTCTACTCCTTCAA-3'	48
CHD281A.R	5'-TTATTTTAGTAGACTGAAAT-3'	
CHD281C.R	5'-TTATTTTAGTAGACTGAAAG-3'	
<i>A. strepera</i>		
CHD171C.F	5'-TTGTTTCCATATCAAATAC-3'	50
CHD171T.F	5'-TTGTTTCCATATCAAATA-3'	
CHD355.R	5'-AGTTCCTCTGCACCAAACCTTA-3'	
CHD1Za		
CHDZa.F	5'-CTGGACATCCTAGCAGAGTACCTA-3'	60
CHDZa.R	5'-TCCTCAATCCCTTTTATTGA-3'	

We also sequenced 391–401 bp of CHD intron a and 36 bp of flanking exon, for a subset of individuals (CHD1Za). We initially amplified this intron using the primers 2550F and 2718R (Ellegren and Fridolfsson, 1997; Fridolfsson and Ellegren, 1999). In females, these primers amplified the intron from the W-chromosome only, but in males, successfully amplified the Z-chromosome. Using these initial sequences, we designed internal primers specific to the Z-chromosome (Table 1). One individual representing each unique allele from CHD1Zb was arbitrarily chosen for additional sequencing. Two *A. strepera* alleles, four *A. penelope* alleles, and two *A. falcata* alleles were not sequenced for CHD1Za because amplification failed or these alleles were sampled only from heterozygotes (the gametic phase of the combined introns could not be determined).

We amplified and sequenced 658–659 bp from domains I and II (5'-end and middle) of the mtDNA control region using the primers L78 and H774 (Sorenson and Fleischer, 1996; Sorenson et al., 1999) and the protocols described above for CHD, except that we used a 52 °C annealing temperature (McCracken et al., 2001). Using the same protocols, we sequenced domain III (3'-end) of the mtDNA control region and part of the phenylalanine tRNA (t-Phe) gene for one arbitrarily chosen representative of each species using the primers L736 and H1251 (Sorenson, unpubl. data; Sorenson et al., 1999). All sequences have been deposited in GenBank (Accession Nos. AY881728–AY881966).

2.3. Phylogenetic inferences

We aligned CHD introns using Sequencher 4.1 (Genecodes, Ann Arbor, MI). For CHD1Zb alleles, only one 1-bp indel was observed, and the alignment was unambiguous. However, several gaps were present in CHD1Za alleles. We systematically shifted gap positions to determine which alignments required the fewest steps (see Sorenson and Payne, 2001), treating multiple-bp gaps as a single step. Ultimately, we coded the presence/

absence of four gaps that varied in size from 2 to 4 bp, and added these codes to the end of the data matrix (Prychitko and Moore, 2000). The lengths of all gaps varied between *Aix sponsa* and other taxa, so we defined three character states for these gaps that corresponded to length; gap length was unordered in parsimony analyses.

Because many of the mtDNA sequences varied in length among and within species due to insertions and deletions of nucleotides, and alignments were ambiguous in several regions, we aligned control region sequences using direct optimization as implemented in POY 2.7 (Gladstein and Wheeler, 2000). We included each unique haplotype (including the eight haplotypes for which we had sequence from all three domains) in this analysis and performed 100 random addition replicates (each limited to five trees), with equal weights for all changes, tree bisection and reconnection (TBR) branch swapping, and an insertion–deletion cost equal to one. Using the implied alignment generated by POY as a guide, we next optimized the alignment for the shortest tree found by POY using methods described by Sorenson and Payne (2001). Next, we systematically shifted the locations of gaps to determine whether combining some gaps as a single evolutionary event decreased the number of steps in the tree. As a result, three gaps were combined into one 8-bp deletion and two gaps were combined into a 2-bp deletion. All gaps were coded as present or absent and added to the end of the data matrix (Prychitko and Moore, 2000). CHD and mtDNA alignments have been placed in TreeBase (study Accession No. S1229).

We next constructed equally weighted maximum parsimony (MP) and maximum likelihood (ML) trees for CHD and mtDNA using PAUP* 4.0b10 (Swofford, 1999). For ML estimates, we chose the most appropriate model of DNA substitution using an arbitrarily chosen MP tree and a hierarchical likelihood ratio test implemented in ModelTest 3.06 (Posada and Crandall, 1998). The initial ML tree was constructed using the chosen model (and parameter estimates) and a different MP

starting tree. Because the number of taxa prohibited direct optimization of parameters, we re-estimated parameters based on the ML tree, and using these parameters, constructed a second ML tree. We repeated this process until tree scores and parameter values stabilized (Wilgenbusch and de Queiroz, 2000). MP and ML trees have been placed in TreeBase (study Accession No. S1229). Statistical support for MP and ML trees was evaluated by bootstrapping (Felsenstein, 1985); we conducted 1000 pseudoreplicates for the combined CHD dataset and the combined mtDNA control region dataset using a heuristic search with the TBR branch-swapping algorithm. For the tree constructed from the 5'-end of control region, we only evaluated bootstrap support for the MP tree by conducting 500 pseudoreplicates because the time required for the analyses was prohibitive.

We compared the combined mtDNA control region (domains I, II, and III) tree and the combined CHD intron (CHD1Za and CHD1Zb) tree to trees in which the topologies were constrained to agree with Livezey's (1991) morphological phylogeny and Johnson and Sorenson's (1998) mtDNA coding region phylogeny (see Fig. 1). Constraint trees were constructed in MacClade 4.0 (Maddison and Maddison, 2000). For CHD, only relationships among species were constrained. Incorporating these constraints, MP and ML trees were constructed using the methods described above. ML trees were compared using a Shimodaira–Hasegawa test with resampling-estimated log likelihood (RELL) bootstrapping and 1000 replicates (SH test, Shimodaira and Hasegawa, 1999). Because ML does not incorporate information from gaps, we also compared MP trees using a two-tailed Templeton test (Templeton, 1983). Topology tests were conducted using PAUP* 4.0b10

(Swofford, 1999). For mtDNA, we only compared relationships within the wigeons and allies, but for CHD, we compared both ingroup relationships, and the relationships between the ingroup and other *Anas* ducks.

To evaluate how much genetic variation was partitioned among and within species, we compared CHD1Zb allele frequencies among wigeons and allies using an analysis of molecular variance (AMOVA, Excoffier et al., 1992; executed in Arlequin 2.0, Schneider et al., 2000). We also calculated pairwise Φ_{ST} statistics among all pairs of species in Arlequin.

3. Results

3.1. CHD—nDNA phylogeny

We sequenced a total of 116 CHD1Zb alleles within the wigeons and allies and found 26 unique alleles, only one of which was shared between multiple species (Fig. 3). Otherwise, alleles clustered into species-specific groupings (Fig. 3). Overall, 71.9% of the total genetic variation was explained by differences among the five species, and these differences were highly significant (AMOVA, $p < 0.0001$). Pairwise comparisons of Φ_{ST} indicated that differences were significant among all species pairs (Φ_{ST} range = 0.50–0.87, $p < 0.0001$). Although CHD1Zb revealed species-specific differences, relationships among species were not well-resolved. To increase the number of phylogenetically informative sites, we sequenced CHD1Za for 19 of the CHD1Zb alleles; one *A. penelope* was heterozygous for CHD1Za, and therefore we included 20 unique alleles in phylogenetic analyses. Combining CHD datasets for a total of 776 characters (including four coded indels; see Section 2),

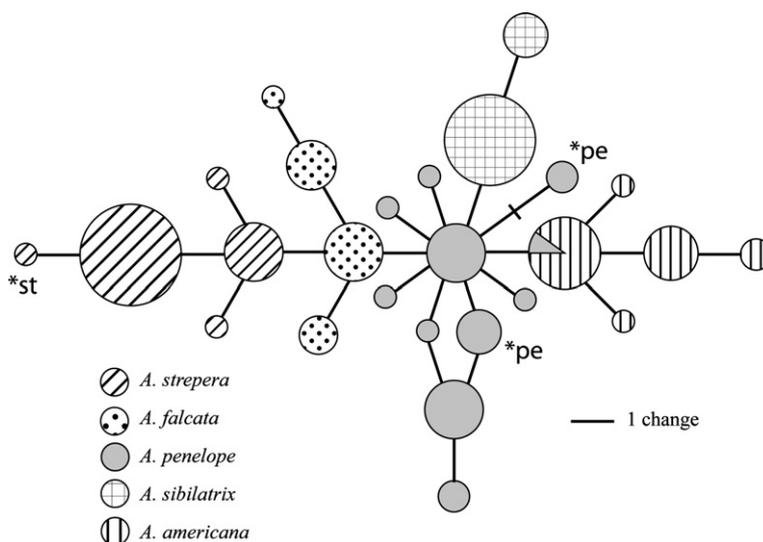


Fig. 3. Unrooted parsimony network showing the relationships among CHD1Zb alleles sampled in the five wigeon species. The areas of circles are proportional to the number of alleles sampled. *st and *pe's indicate those alleles that were sampled from *A. strepera* and *A. penelope*, respectively, that had mtDNA that disagreed with species boundaries (see text).

we found 70 (9.0%) variable sites, 21 (2.7%) of which were parsimony informative, and the combined datasets provided greater phylogenetic signal for among species relationships (Fig. 4). ModelTest chose the Hasegawa, Kishino, and Yano model (HKY; Hasegawa et al., 1985) with unequal base frequencies and a transition to transversion bias (ti/tv = 1.3654).

MP and ML analyses yielded largely congruent trees (see Fig. 4). All wigeon alleles formed a clade with moderate–high bootstrap support (87%), and *A. penelope* was paraphyletic to *A. americana* and *A. sibilatrix* (Fig. 4). Half of the MP trees recovered *A. americana* as monophyletic (Fig. 4A), but the remaining MP trees and the ML tree suggested that *A. americana* is paraphyletic to *A. sibilatrix* and polyphyletic with *A. penelope* (Fig. 4B). In agreement with mtDNA, *A. strepera* and *A. falcata* were recovered as sister species (87 and 65% bootstrap support for MP and ML analyses, respectively). *A. falcata* might be paraphyletic to *A. strepera*, but their relationship is also consistent with reciprocal monophyly (Fig. 4). Also agreeing with mtDNA, the wigeons and allies grouped with *A. acuta* and *A. carolinensis* to

the exclusion of *A. clypeata* and *A. discors* (the blue-winged ducks), but this relationship was not strongly supported (63 and 62% bootstrap support for MP and ML analyses, respectively). CHD did not recover a monophyletic wigeons and allies clade, although our data also could not refute this possibility.

For maximum likelihood, the CHD nuclear intron tree did not differ significantly from trees constrained to agree with the topologies of the mtDNA or morphology trees (Table 2). However, incorporating information from gaps in MP analyses indicated that the CHD tree differed marginally from the tree constrained to agree with mtDNA, but differed significantly from one constrained to agree with morphology (Table 2). Some of this disagreement can be explained by the strongly supported paraphyly of *A. penelope*. For example, allowing *A. penelope* to be paraphyletic to *A. sibilatrix* and *A. americana*, but forcing a sister-relationship between *A. sibilatrix* and *A. americana*, improved the fit between CHD and mtDNA in both ML and MP comparisons (Table 2). However, allowing *A. penelope* to be paraphyletic to *A. americana* (i.e., forcing a monophyletic

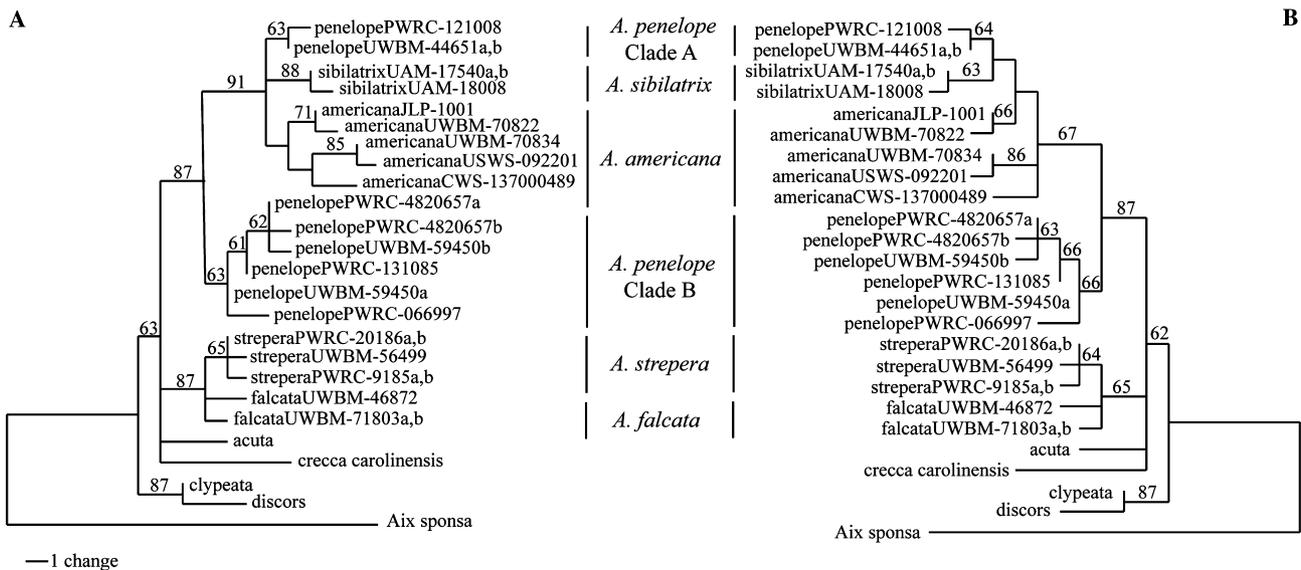


Fig. 4. Two of the six equally parsimonious trees (length = 76 steps, CI = 0.987) constructed from the combined CHD introns (CHD1Za and CHD1Zb) rooted with *A. sponsa*. Half of the MP trees recovered a monophyletic *A. americana* (A), whereas the remaining trees and the ML tree ($-\ln L = 1523.37$) recovered a paraphyletic *A. americana* (B; the ML topology was identical to this tree). MP (A) and ML (B) bootstrap support that was greater than 50% is shown above branches. Alleles isolated from males, for which the gametic phase of the combined introns could be determined (i.e., they were homozygous for at least one intron), are indicated as alleles a and b.

Table 2
Comparisons between the CHD intron phylogeny and trees constrained to agree with mtDNA and morphology trees

Topological constraint	ML–SH test		MP–Templeton test		
	ln L difference	p	Length difference	z	p
mtDNA	9.84	0.26	3	1.73	0.08
<i>A. penelope</i> paraphyletic	0.65	0.82	0	0.0	1.0
Morphology	18.31	0.14	6	2.44	0.01
<i>A. penelope</i> paraphyletic	18.31	0.14	6	2.44	0.01

“northern wigeon,” *sensu* Livezey, 1991), but not *A. sibilatrix*, did not improve the fit between CHD and morphology (Table 2). Clearly, the CHD tree was more congruent with mtDNA than it was with morphology.

3.2. mtDNA phylogeny

The size of the mtDNA control region sequence varied from 658 to 659 bp within the wigeons and allies and yielded an alignment of 660 positions. Outgroup sequences ranged from 658 to 667 bp, and the alignment including the ingroup and outgroup was 676 bp in length. There were 20 gaps in our alignment, which we coded as present or absent, resulting in a total of 696 characters, of which 125 (18.0%) were variable and 73 (10.5%) were parsimony informative. The best-fit ML model selected using ModelTest was the Tamura Nei (TrN; Tamura and Nei, 1993) model with invariable sites (I) and a gamma distribution (Γ), which includes parameters for unequal base frequencies, uneven ratios of C–T and A–G transitions, invariant sites, and unequal rates of nucleotide substitution among sites.

Most haplotypes from each species grouped into species-specific monophyletic groups with moderate to high bootstrap support (75–99%, Fig. 5), but three examples of mtDNA non-monophyly were observed. Two *A. penelope* shared haplotypes with *A. americana*, and were grouped within a clade containing all of the *A. americana* haplotypes (Fig. 5). One *A. strepera* shared a haplotype with three *A. penelope*, and this haplotype grouped within a clade containing the majority of the *A. penelope* haplotypes. These examples of paraphyly/polyphyly were observed in 100% of MP trees and the ML tree (not shown), and received moderate to high bootstrap support in parsimony analyses (75–93%, Fig. 5). All three individuals that had mtDNA disagreeing with species boundaries had CHD alleles that clustered with other conspecifics (Fig. 3; Appendix). These examples of haplotype sharing likely resulted from interspecific hybridization rather than deep cryptic lineages or incomplete sorting of ancestral polymorphisms (see Section 4 below).

Ignoring the putative introgressed haplotypes, there was moderate to strong support for the monophyly of each species, but the relationships among the species were not well-resolved. The ML tree ($-\ln L = 1966.5$) and 100% of MP trees agreed in placing *A. sibilatrix* sister to *A. americana*, but this relationship was not strongly supported (54% MP bootstrap support; Fig. 5). *A. strepera* and *A. falcata* formed a sister-group in all our analyses, and this relationship received 100% MP bootstrap support. The relationship between *A. penelope* and the other species, however, was not well-supported (<50% MP bootstrap support). Ninety-eight percent of MP trees were consistent with mtDNA coding regions suggesting that *A. penelope* is sister to the *americana*–

sibilatrix clade (Johnson and Sorenson, 1998, 1999). Our ML analysis suggested a sister-relationship between *A. penelope* and the *strepera*–*falcata* clade (not shown). The ML tree was only 1-step longer (length = 191 steps) than MP trees that did not include gaps.

The combined mtDNA control region sequences (3′-, middle, and 5′-end) varied between 981 and 982 bp for the ingroup taxa. We excluded 23 bp of t-Phe sequence so that ingroup and outgroup sequences were completely homologous. Truncated ingroup and outgroup sequences yielded a 983 bp alignment. After adding character codes for 29 gaps, the combined control region contained 1011 characters, 166 (16.4%) of which were variable and 75 (7.4%) were parsimony informative. The best-fit model selected by ModelTest was the TrN+I (Tamura and Nei, 1993) model of evolution, which allows for unequal base frequencies, uneven ratios of C–T and A–G transitions, and invariant sites. This larger segment of control region provided additional phylogenetic signal, and this dataset better resolved the relationships within the wigeons and allies. For this dataset, two equally parsimonious trees were found that disagreed only in the relationships among the outgroup taxa, and one of these trees was identical to the ML tree ($-\ln L = 2330.5$; Fig. 6). The sister taxon relationship between *A. americana* and *A. sibilatrix* was strongly supported in both MP and ML analyses (98 and 97% bootstrap support, respectively), and the placement of *A. penelope* as sister to the *americana*–*sibilatrix* clade instead of the *strepera*–*falcata* clade was moderately supported (76% bootstrap support for both MP and ML). The monophyly of the wigeons and allies also was supported by high bootstrap values (88 and 97% for MP and ML analyses, respectively). The relationships among the ingroup taxa are identical to the mtDNA coding region phylogeny of Johnson and Sorenson (1998, 1999). Constraining the tree topology to agree with Livezey’s (1991) morphology tree required 26 additional steps (i.e., 253 steps in total), a significant difference (Templeton test, $z = 4.75$, $p < 0.0001$), and this tree also received a significantly worse ML-score (SH test, $\ln L$ difference = 40.25, $p = 0.001$).

4. Discussion

4.1. nDNA phylogeny—CHD

Relative to other nuclear introns, CHD introns showed remarkable utility as species-level phylogenetic markers. Within the wigeons and allies, we found 26 CHD1Zb alleles, and only one of these was shared between species (Fig. 3). Furthermore, alleles tended to cluster into species-specific groupings, and 72% of the total genetic variation was explained by differences among species. Combining CHD1Zb and CHD1Za

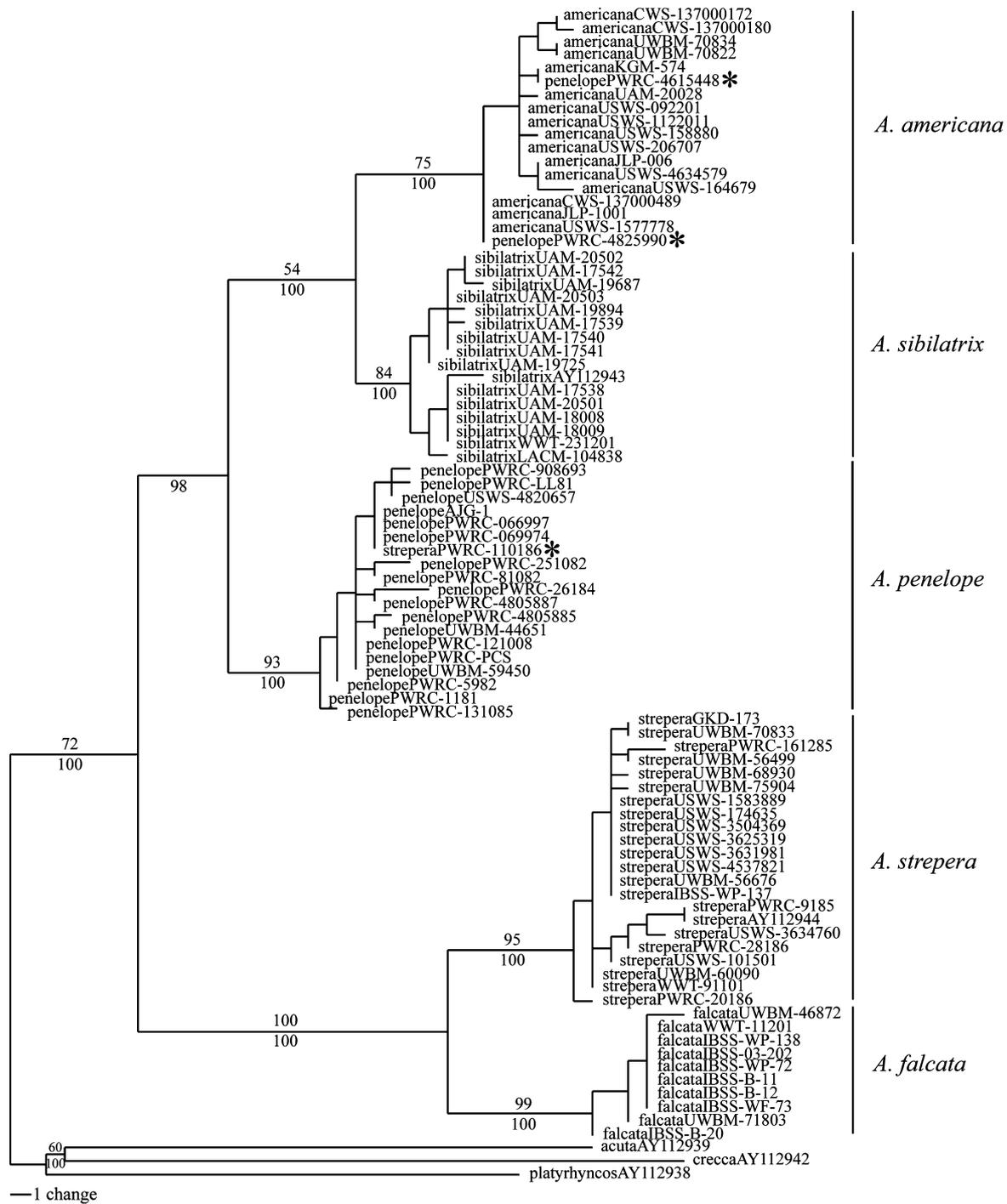


Fig. 5. Mitochondrial DNA control region phylogeny (domains I and II). MP found more than 150,000 equally parsimonious trees (length = 209 steps, CI = 0.649). MP bootstrap support is shown above branches (>50%); frequency values for nodes supported by the 50% majority-rule consensus tree are shown below branches. Values are shown for deeper nodes only. Individuals that have mtDNA that disagrees with species boundaries (i.e., putative hybrids) are indicated by asterisks (*). The ML topology was similar to this tree, except that *A. penelope* was sister to the *strepera*–*falcata* clade and relationships among individuals within taxa varied.

provided strong phylogenetic signal: four of the five species were consistent with monophyletic clades, several nodes were supported by moderate to high bootstrap support, and MP trees received a high consistency index (CI = 0.986, Fig. 4). In agreement with both mtDNA (Johnson and Sorenson, 1998, 1999) and morphology

(Livezey, 1991), CHD resolved a monophyletic group consisting of the three wigeon species, and this node received high bootstrap support in both MP and ML analyses (Fig. 4). However, CHD did not recover a monophyletic clade consisting of the five wigeons and allies, nor a sister-relationship between *A. crecca* and *A.*

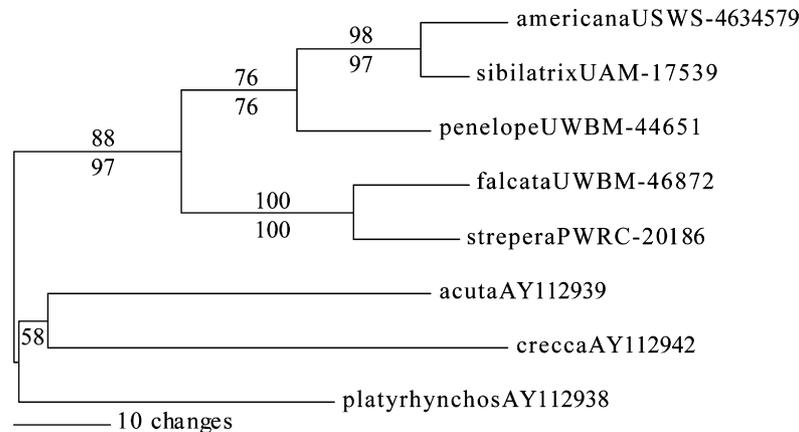


Fig. 6. Mitochondrial DNA control region phylogeny (domains I, II, and III). MP found two equally parsimonious that only differed in the relationships among outgroup taxa (length = 227 steps, CI = 0.79). The topology of the ML tree was identical to the tree shown here ($-\ln L = 2330.5$). MP bootstrap values that were greater than 50% are shown above each branch; ML bootstrap values are shown below each branch.

acuta. Both of these relationships were supported by morphology and mtDNA. In addition, CHD did not resolve the relationships among the three wigeon species, because *A. penelope* was paraphyletic and support for relationships within the clade shared by the three wigeon species was low (see Fig. 4). The multi-furcating CHD tree suggests that its resolving power was low in some cases, but this power might be enhanced by sequencing additional linked introns. Sundstrom et al. (2003) sequenced five different CHD introns, and combining several of these introns could offer more characters for constructing a well-resolved gene tree.

The surprising observation that CHD alleles in most species have sorted to monophyly contrasts markedly with nuclear intron phylogenies for other groups of closely related species. Allen and Omland (2003) evaluated ornithine decarboxylase (ODC) among closely related species of the genus *Icterus* (New World orioles). Although ODC recovered the same three deeply divergent clades identified by mtDNA (see Omland et al., 1999), relationships among species within these clades were not well-resolved, and several species shared alleles or were polyphyletic. Johnson and Clayton (2000a) similarly found shared alleles and polyphyly among closely related *Zenaida* species (New World doves) using β -fibrinogen intron 7 (FIB7). The strong phylogenetic signal provided by CHD (sex-linked introns), relative to ODC and FIB7 (autosomal loci), may be attributed to the lower effective population size of Z-chromosome-linked loci, and the resulting slightly more rapid sorting rates. However, Pitra et al. (2002) found weak phylogenetic signal from homologous CHD introns, leading them to exclude CHD from phylogenetic analyses. Comparing our results with those of Pitra et al. (2002) suggests that the phylogenetic utility of CHD found in this study might not be characteristic of Z-linked-loci, but may be specific to the wigeons and allies.

The strong phylogenetic signal provided by CHD could be explained if the wigeons and allies experienced recent bottlenecks. Founder events leading to peripheral isolation (Mayr, 1942) or post-speciation bottlenecks could cause nuclear loci to sort much more rapidly. Johnson and Sorenson (1999), for example, found little biogeographic signal using their mtDNA phylogeny of *Anas* species, and they suggested that widespread dispersal in ducks may have promoted dispersal-driven divergence (i.e., divergence via peripheral isolation). When divergence is driven by dispersal and peripheral isolation, the original founders will carry a subset of the ancestral allelic diversity to the founded area, and genetic drift may eliminate additional alleles as a result of the subsequent small population size. Hence, dispersal-driven divergence can cause alleles to sort to monophyly rapidly. The tight clustering of species-specific CHD alleles within the wigeons and allies is expected under Johnson and Sorenson's (1999) dispersal-driven divergence hypothesis. Alternatively, strong selection on coding regions of CHD or other closely linked genes could cause these introns to sweep through the population, and hence, differences among species could become fixed rapidly (Donnelly and Tavaré, 1995). Sequencing additional nuclear loci will be useful for differentiating between these hypotheses (Galtier et al., 2000).

4.2. Sampling multiple individuals—paraphyly/polyphyly

Although we did not find deep cryptic lineages within species, dense sampling of each species revealed paraphyly and polyphyly within the wigeons and allies for both mtDNA and CHD. Specifically, two *A. penelope* shared mtDNA haplotypes with *A. americana*, and one *A. strepera* shared a mtDNA haplotype with three *A. penelope* (Fig. 5). These haplotypes were nested within

well-supported clades containing all of the *A. americana* haplotypes and the majority of *A. penelope* haplotypes, respectively. In addition, dense sampling of individuals revealed a CHD1Zb allele that was shared between *A. americana* and *A. penelope* (Fig. 3), and the combined CHD dataset clearly showed *A. penelope* to be paraphyletic to the other wigeons (Fig. 4).

Mitochondrial DNA haplotype sharing in this group probably resulted from interspecific hybridization. Compared to the shallow differences observed within clades, deeply divergent lineages separated the major clades of all species, suggesting long periods of isolation and divergence. In addition, *Anas* ducks are well known for their capacity to hybridize and produce fertile offspring (Johnsgard, 1960; Tubaro and Lijtmaer, 2002). *A. americana* and *A. penelope* regularly hybridize in the wild (Johnsgard, 1965; Mowbray, 1999; Sonobe and Usui, 1993), and *A. penelope* × *A. strepera* hybrids have also been observed (Johnsgard, 1965; LeSchack et al., 1997). The two *A. penelope* that shared mtDNA haplotypes with *A. americana* were both sampled from the west coast of North America where the two species' wintering distributions overlap (see Edgell, 1984). These two species also regularly come into contact in Eurasia during the winter (Cramp et al., 1977; Peterson et al., 1983; Sonobe and Usui, 1993). In waterfowl, pair formation occurs on the wintering grounds (Rohwer and Anderson, 1988), and range overlap in winter might facilitate hybridization.

In contrast to mtDNA, the paraphyly of *A. penelope* observed in CHD probably is better explained by ancestral polymorphism. CHD has three times the effective population size of mtDNA, and sorting to monophyly will occur much more slowly (Hudson and Turelli, 2003; Palumbi et al., 2001). Hence, ancestral polymorphisms will be retained in the population long after mtDNA has sorted to monophyly. The clustering of *A. penelope* CHD1Zb alleles, the independent derivation of *A. americana* and *A. sibilatrix* alleles from an "ancestral" *A. penelope* allele (Fig. 3; see Castelleo and Templeton, 1994), and two distinct lineages of *A. penelope* in the combined CHD introns phylogeny (Fig. 4) further argues for ancestral polymorphism. Although ancestral polymorphism seems to be the better explanation of paraphyly in CHD, the possibility of interspecific hybridization cannot be ruled out.

In this study, the level of interspecific mtDNA haplotype sharing was low, but it does reinforce the point that using only one representative of each species can mislead phylogenetic inferences (Funk and Omland, 2003; Omland, 1997; Omland et al., 1999). At least in this group of ducks, two representatives would usually be sufficient to improve confidence in phylogenetic relationships, because the probability of sampling two introgressed haplotypes is low. In contrast, Kulikova et al. (2004) sampled 22 individuals of *Anas zonorhyncha* (eastern

spot-billed duck) and found two common clades of mtDNA haplotypes. Kulikova et al. (2004) concluded that the single haplotype sequenced by Johnson and Sorenson (1998, 1999) resulted from introgression, and that the previously unidentified clade originally belonged to *A. zonorhyncha*. These results have challenged former inferences of phylogenetic relationships among mallard species. Similarly, in this study, one representative of CHD would not have revealed the paraphyly of *A. penelope*, and conclusions regarding the relationships among the three wigeon species would vary depending on which clade of *A. penelope* was sampled. Sampling multiple individuals of each taxon for phylogenetic analyses will be most important when interspecific hybridization is common and when linkage groups, especially nuclear introns, may have had an insufficient amount of time to sort to monophyly (see Funk and Omland, 2003).

4.3. DNA, morphology, and biogeography

In this study, phylogenies constructed from DNA and morphology are largely incongruent (see also Johnson and Sorenson, 1999), whereas the mtDNA and CHD trees are congruent. mtDNA recovered *A. americana* as sister to *A. sibilatrix*, with this relationship receiving high bootstrap support from both coding regions and the combined control region datasets (100 and 98%, respectively; Johnson and Sorenson, 1998, 1999; Fig. 6, this study). In contrast, all 157 morphological characters scored by Livezey (1991) were shared between *A. americana* and *A. penelope*, whereas *A. sibilatrix* differed from these two species by seven characters, six of which were shared with *A. strepera* and *A. falcata*. Behavioral displays, including courtship displays and family rituals, also indicate that *A. americana* is phenotypically more similar to *A. penelope* than to *A. sibilatrix* (Brewer, 1997; Johnsgard, 1965). Although CHD was not conclusive, the basal position of an exclusively *A. penelope* clade, and the ancestral position of an *A. penelope* CHD1Zb allele, strongly argues against an earlier divergence of *A. sibilatrix*, but argues for a recent derivation of *A. sibilatrix* from a northern ancestor (also see Johnson and Sorenson, 1999; Livezey, 1991). These observations raise an interesting question; why is there such profound disagreement between phenotype and genotype among the wigeon species?

Interestingly, a second clade of *Anas* ducks has a biogeographic pattern and a conflict between mtDNA and morphology that is analogous to that found in the wigeons. The green-wing teal (*Anas crecca*) includes two subspecies from the northern hemisphere, *A. c. crecca* (Eurasian common teal) and *A. c. carolinensis* (North American green-winged teal) that are almost identical morphologically (Livezey, 1991). However, mtDNA indicates that *A. c. carolinensis* is sister to the South

American species-complex, *A. flavirostris* (speckled teal; Johnson and Sorenson, 1998, 1999), but these taxa differ substantially in morphology (Livezey, 1991). These congruent patterns between wigeons and green-winged teal possibly resulted from selective pressures that differed between southern and northern hemispheres and caused rapid divergence in phenotypic characters following the colonization of South America from North America (see Johnson, 1999; Johnson et al., 1999). Hence, phenotypes may have accumulated diagnostic differences much more rapidly than DNA (see also Cunningham et al., 1992; Kondo et al., 2004). The parallel discordance between morphology and DNA in the wigeons and the green-winged teal offers an exciting opportunity to investigate potential interactions between peripheral isolation and disruptive selection during speciation, using comparative phylogeography.

There also exists disagreement between phenotype and DNA in the placement of *A. falcata*. Morphology and behavior both suggest that *A. falcata* is an evolutionary link between the wigeon species and *A. strepera*, whereas *A. strepera* is intermediate between the wigeon species (including *A. falcata*) and other *Anas* species (Johnsgard, 1965; Livezey, 1991). However, mtDNA strongly favors *A. falcata* as sister to *A. strepera*, not the wigeon species (Johnson and Sorenson, 1998, 1999; Figs. 5 and 6, this study), and CHD corroborated this relationship (Fig. 4). Again, why is there strong disagreement between phenotype and genotype? It seems likely that some of this disagreement within *Anas* ducks might be explained by convergence or reversals in phenotypic characters (e.g., McCracken et al., 1999; Omland and Lanyon, 2000), but ancient introgression of DNA from close relatives also might have played a role (Ballard and Whitlock, 2004).

5. Conclusions

We found CHD introns to have strong phylogenetic potential compared to other nuclear introns. However, it is unclear whether this phylogenetic signal is best explained by: (1) the hypothesized faster sorting rate of Z-linked loci relative to autosomal loci, (2) extreme bottlenecks in this group of ducks, perhaps coincident with dispersal-driven divergence, or (3) strong selection on closely linked loci. Regardless, the phylogenetic utility of CHD introns should be evaluated in other taxa for

which a well-resolved gene tree is desired. Also, within this group of ducks, there is strong disagreement between phenotype and DNA in the relationships among the wigeons and allies, but the evolutionary implications of this conflict are unclear. To examine this conflict in more detail, a comparison between morphology, mtDNA, and additional nuclear DNA is needed.

Acknowledgments

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Appendix

Collection information and identification of CHD alleles for all individuals of wigeons and allies included in this study

Species	Identification number ^a	Country: state or province	Season ^b	CHD alleles ^{c,d}	Tissue type
<i>Anas strepera</i>	PWRC-161285	Netherlands	NB	st2, st2	Muscle
<i>A. strepera</i>	PWRC-9185	Netherlands	NB	st2, st2	Muscle
<i>A. strepera</i>	PWRC-20186	Netherlands	NB	st3, st3	Muscle
<i>A. strepera</i>	PWRC-28186	Netherlands	NB	?	Muscle
<i>A. strepera</i>	PWRC-110186	Netherlands	NB	st1	Muscle
<i>A. strepera</i>	UWBM-56499	Russia: Astrakhanskaya Oblast [†]	B	st4	Muscle
<i>A. strepera</i>	UWBM-56676	Russia: Astrakhanskaya Oblast [†]	B	st1, st1	Muscle
<i>A. strepera</i>	UWBM-60090	Mongolia: Donrod Aymag	B	st3	Muscle
<i>A. strepera</i>	UWBM-75904	Russia: Respublika Tyva	B	st1, st5	Muscle
<i>A. strepera</i>	IBSS-WP-137	Russia: Primorskiy Kray	NB	st3, st3	Muscle
<i>A. strepera</i>	WWT-91101	United Kingdom	NB	st2, st2	Feathers
<i>A. strepera</i>	UWBM-68930	USA: California	B	st2, st2	Muscle
<i>A. strepera</i>	GKD-173	USA: Washington	NB	st2	Muscle
<i>A. strepera</i>	UWBM-70833	USA: Utah	B	st2	Muscle
<i>A. strepera</i>	USWS-3631981	USA: North Dakota	B	st2, st2	Muscle
<i>A. strepera</i>	USWS-3504369	USA: Colorado	B	st2	Muscle
<i>A. strepera</i>	USWS-4537821	USA: Nevada	B	st2, st2	Muscle
<i>A. strepera</i>	USWS-1746935	USA: New York	B	st2	Muscle
<i>A. strepera</i>	USWS-3625319	USA: South Dakota	B	st2	Muscle
<i>A. strepera</i>	USWS-1583889	USA: Wyoming	B	st2, st2	Muscle
<i>A. strepera</i>	USWS-101501	USA: Alaska	B	st3	Muscle
<i>A. strepera</i>	USWS-3634760	USA: Montana	B	st2, st3	Muscle
<i>A. falcata</i>	UWBM-71803	Russia: Primorskiy Kray	B	fa3, fa3	Muscle
<i>A. falcata</i>	UWBM-46872	Russia: Khabarovkiy Kray	B	fa1	Muscle
<i>A. falcata</i>	WWT-11201	United Kingdom: captivity	NA	fa1	Feathers
<i>A. falcata</i>	IBSS-B-20	Russia: Primorskiy Kray	NB	fa3, fa3	Muscle
<i>A. falcata</i>	IBSS-WP-138	Russia: Primorskiy Kray	NB	fa3	Muscle
<i>A. falcata</i>	IBSS-03-202	Russia: Primorskiy Kray	NB	fa1, fa2	Muscle
<i>A. falcata</i>	IBSS-WP-72	Russia: Primorskiy Kray	NB	fa1	Muscle
<i>A. falcata</i>	IBSS-B-11	Russia: Primorskiy Kray	NB	fa2, fa4	Muscle
<i>A. falcata</i>	IBSS-B-12	Russia: Primorskiy Kray	NB	fa1, fa1	Muscle
<i>A. falcata</i>	IBSS-WF-73	Russia: Primorskiy Kray	NB	fa2, fa2	Muscle
<i>A. sibilatrix</i>	UAM-18008	East Falkland Islands	B	si2	Blood
<i>A. sibilatrix</i>	UAM-18009	East Falkland Islands	B	si2	Blood
<i>A. sibilatrix</i>	UAM-17540	Argentina: Rio Negro	B	si1, si1	Muscle
<i>A. sibilatrix</i>	UAM-17539	Argentina: Rio Negro	B	si1, si1	Muscle
<i>A. sibilatrix</i>	UAM-17542	Argentina: Rio Negro	B	si1, si1	Muscle
<i>A. sibilatrix</i>	UAM-17541	Argentina: Rio Negro	B	si2	Muscle
<i>A. sibilatrix</i>	UAM-17538	Argentina: Rio Negro	B	si1	Muscle
<i>A. sibilatrix</i>	UAM-20501	Argentina: Neuquen	B	si1	Muscle
<i>A. sibilatrix</i>	UAM-20502	Argentina: Neuquen	B	si1, si1	Muscle
<i>A. sibilatrix</i>	LACM-104838	Argentina: Neuquen	B	si1, si1	Muscle
<i>A. sibilatrix</i>	UAM-20503	Argentina: Chubut	B	si1	Muscle
<i>A. sibilatrix</i>	UAM-19894	Argentina: Chubut	B	si1	Muscle
<i>A. sibilatrix</i>	UAM-19687	Argentina: Santa Cruz	B	si2	Muscle
<i>A. sibilatrix</i>	UAM-19725	Argentina: Santa Cruz	B	si1	Muscle
<i>A. sibilatrix</i>	WWT-231201	United Kingdom: captivity	NA	si1, si1	Feathers
<i>A. americana</i>	UAM-20028	USA: Alaska	NB	ampe1, am3	Muscle
<i>A. americana</i>	KGM-574	USA: Alaska	B	ampe1	Muscle
<i>A. americana</i>	UWBM-70822	USA: Utah	B	am4	Muscle
<i>A. americana</i>	UWBM-70834	USA: Utah	B	ampe1	Muscle
<i>A. americana</i>	CWS-137000172	Canada: Quebec	B	ampe1	Muscle
<i>A. americana</i>	CWS-137000180	Canada: Quebec	B	ampe1, am3	Muscle
<i>A. americana</i>	CWS-137000489	Canada: Alberta	B	am1	Muscle
<i>A. americana</i>	JLP-006	USA: North Dakota	B	am4	Foot-webbing
<i>A. americana</i>	USWS-1122011	USA: Maine	NB	ampe1, am3	Muscle
<i>A. americana</i>	USWS-092201	USA: Alaska	NB	am2	Muscle
<i>A. americana</i>	USWS-206707	USA: North Dakota	B	ampe1, ampe1	Muscle
<i>A. americana</i>	USWS-158880	USA: Alaska	NB	ampe1	Muscle
<i>A. americana</i>	USWS-164679	USA: Alaska	NB	am3	Muscle

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Appendix (continued)

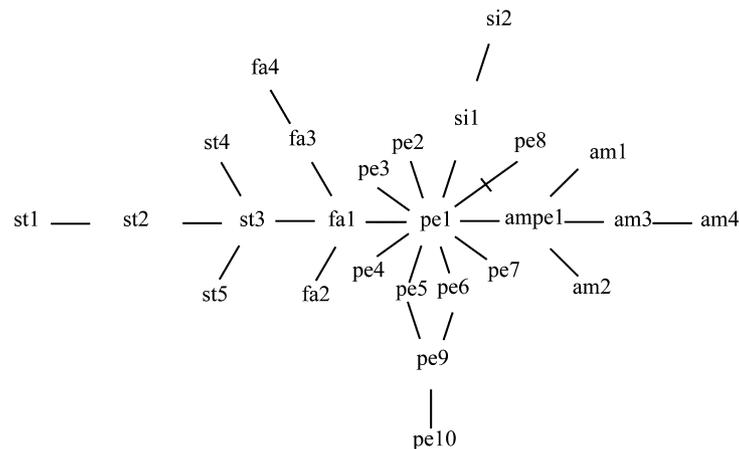
Species	Identification number ^a	Country: state or province	Season ^b	CHD alleles ^{c,d}	Tissue type
<i>A. americana</i>	USWS-157778	USA: New York	B	am3	Muscle
<i>A. americana</i>	JLP-1001	USA: North Dakota	B	am3	Foot-webbing
<i>A. americana</i>	USWS-4634579	USA: Alaska	B	?	Muscle
<i>A. penelope</i>	UWBM-59450	Russia: Yamalo-Nenetskiy Avtonomnyy Okrug	B	pe1, pe10	Muscle
<i>A. penelope</i>	UWBM-44651	Russia: Magadanskaya Oblast	B	pe1, pe1	Muscle
<i>A. penelope</i>	PWRC-LL81	United Kingdom: Scotland	NB	pe5	Muscle
<i>A. penelope</i>	PWRC-1181	United Kingdom: Scotland	NB	pe1, pe8	Muscle
<i>A. penelope</i>	PWRC-251082	Denmark	NB	pe3, pe9	Muscle
<i>A. penelope</i>	PWRC-26184	Denmark	NB	pe9, pe10	Muscle
<i>A. penelope</i>	PWRC-81082	Denmark	NB	pe9	Muscle
<i>A. penelope</i>	PWRC-5982	Denmark	NB	pe1	Muscle
<i>A. penelope</i>	PWRC-131085	Denmark	NB	pe6	Muscle
<i>A. penelope</i>	AJG-1	Spain	B	pe6	Intestine
<i>A. penelope</i>	PWRC-4820657	USA: Washington	NB	pe9, pe9	Muscle
<i>A. penelope</i>	PWRC-908693	USA: Washington	NB	pe1, pe2	Muscle
<i>A. penelope</i>	PWRC-066997	USA: Oregon	NB	pe8	Muscle
<i>A. penelope</i>	PWRC-069974	USA: Oregon	NB	pe1	Muscle
<i>A. penelope</i>	PWRC-4615448	USA: California	NB	pe6	Muscle
<i>A. penelope</i>	PWRC-121008	USA: California	NB	pe7	Muscle
<i>A. penelope</i>	PWRC-4825990	USA: Idaho	NB	pe8, pe10	Muscle
<i>A. penelope</i>	PWRC-PCS	USA: Massachusetts	NB	pe9	Muscle
<i>A. penelope</i>	PWRC-4805885	USA: Alaska	NB	ampe1, pe9	Muscle
<i>A. penelope</i>	PWRC-4805887	USA: Alaska	NB	pe4, pe6	Muscle

^a Museums are abbreviated as follows for voucher specimens: UWBM, University of Washington Burke Museum; UAM, University of Alaska Museum; LACM, Natural History Museum of Los Angeles County; IBSS (Institute of Biology and Soil Sciences, Russian Academy of Sciences); KGM, collected by K.G. McCracken (UAF); GKD, collected by G.K. Davis (UWBM). Abbreviations for nonvouchered specimens are as follows: PWRC, Patuxent Wildlife Research Center; USWS, United States Wing Survey; WWT, Wildfowl & Wetlands Trust; CWS, Canadian Wing Survey; AJG, collected by A.J. Green; JLP, collected by J.L. Peters.

^b Season in which individuals were sampled. B, breeding season; NB, non-breeding season; and NA, not applicable (sampled in captivity).

^c Two CHD alleles indicate a male individual, one allele indicates a female, and a ? indicates that CHD was not sequenced for that individual.

^d Key to CHD alleles (also see Fig. 3).



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